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(54) Title: AFAMIN: A HUMAN SERUM ALBUMIN-LIKE PROTEIN

(57) Abstract

The invention relates to a novel human serum protein referred to as AFM, which has one or more activities in common with human serum albumin, human α -fetoprotein, or human vitamin D binding protein and which has an apparent molecular weight by SDS-PAGE of 87 kd; variants thereof; and related genes, vectors, cells and methods.

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AFAMIN: A HUMAN SERUM ALBUMIN-LIKE PROTEIN

Field of the Invention

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Generally, the invention relates to the field of human serum proteins that are functionally and structurally similar to the related proteins: human serum albumin (ALB), human α -fetoprotein (AFP), and 10 vitamin D-binding protein (VDB).

Background of the Invention

The human serum proteins albumin (ALB), 15 α -feto-protein (AFP) and vitamin D binding protein (VDB) are known to be members of a multigene ALB family. All three proteins are found in serum where they mediate the transport of a wide variety of ligands. ALB binds fatty acids, amino acids, steroids, glutathione, metals, 20 bilirubin, lyssolecithin, hematin, prostaglandins and pharmaceuticals (for review, see 1). AFP binds fatty acids, bilirubin and metals (2, 3). VDB binds vitamin D and its metabolites as well as fatty acids, actin, C5a and C5a des Arg. (4-7).

25 In addition to their transport capabilities, ALB family proteins possess a wide assortment of other functional activities. ALB is the main contributor to the colloid oncotic pressure of plasma, acts as a scavenger of oxygen-free radicals and can inhibit 30 copper-stimulated lipid peroxidation, hydrogen peroxide release, and neutrophil spreading (1, 8-10). AFP has been implicated in the regulation of immune processes (11-14) and VDB can act as a co-chemotactic factor for neutrophils (6, 15) and as an activating factor for 35 macrophages (16).

The serum levels of ALB family proteins are also known to be responsive to various pathological conditions. ALB is a negative acute phase protein (17) whose levels decrease in times of stress. AFP levels are 5 elevated in women carrying fetuses with certain developmental disorders (18, 19) and in individuals with hepatocarcinoma, teratocarcinoma, hereditary tyrosinemia or ataxia-telangiectasia (20-24). VDB levels are decreased in patients with septic shock (25) or 10 fulminant hepatic necrosis (26, 27).

ALB family members also have significant structural similarities. Homology has been observed at the primary amino acid sequence level and there is also a well-conserved pattern of Cys residues which predicts 15 similar secondary structures (28-32). ALB family genes have similar exon/intron organizations (33-36) and all have been mapped to human chromosome 4 within the region 4q11-q22 (37, 38).

Human "Afamin" (abbreviated as "AFM") is a 20 novel serum protein with a molecular weight of 87000 daltons. It shares strong similarity to albumin family members and has the characteristic pattern of disulfide bonds observed in this family. In addition, the gene maps to chromosome 4 as do other members of the albumin 25 gene family. Thus, AFM is the fourth member of the albumin family of proteins. AFM cDNA was stably transfected into Chinese hamster ovary cells and recombinant protein (rAFM) was purified from conditioned medium. Both rAFM and AFM purified from human serum 30 react with a polyclonal antibody that was raised against a synthetic peptide derived from the deduced amino acid sequence of AFM. It is expected that AFM will have properties and biological activities in common with ALB, AFP, and VDB.

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25 The sections below contain a summary of background information that is currently available on ALB, AFP, and VDB and contains lists of additional publications relating to these known proteins.

30 I. Human Serum Albumin

Human serum albumin is an important factor in the regulation of plasma volume and tissue fluid balance through its contribution to the colloid osmotic pressure of plasma. Albumin normally constitutes 50-60% of

plasma proteins and because of its relatively low molecular weight (66,300-69,000), exerts 80-85% of the colloidal osmotic pressure of the blood.

The best known functions of ALB involve regulation of transvascular fluid flux and hence, intra and extravascular fluid volumes and transport of lipid and lipid-soluble substances. ALB solutions are frequently used for plasma volume expansion and maintenance of cardiac output in the treatment of certain types of shock or impending shock including those resulting from burns, surgery, hemorrhage, or other trauma or conditions in which a circulatory volume deficit is present. Transfusions of whole blood or red blood cells also may be necessary, depending on the severity of red blood cell loss.

Intravenous (IV) administration of concentrated ALB solutions causes a shift of fluid from the interstitial spaces into the circulation and a slight increase in the concentration of plasma proteins. When administered IV to a well-hydrated patient, each volume of 25% ALB solution draws about 3.5 volumes of additional fluid into the circulation within 15 minutes, reducing hemoconcentration and blood viscosity. In patients with reduced circulating blood volumes (as from hemorrhage or loss of fluid through exudates or into extravascular spaces), hemodilution persists for many hours, but in patients with normal blood volume, excess fluid and protein are lost from the circulation within a few hours. In dehydrated patients, ALB generally produces little or no clinical improvement unless additional fluids are administered.

Although ALB contains some bound amino acids, it provides only modest nutritive effect. ALB binds and functions as a carrier of intermediate metabolites (including bilirubin), trace metals, some drugs, dyes, fatty acids, hormones, and enzymes, thus affecting the

transport, inactivation, and/or exchange of tissue products.

ALB is also involved in a number of other vital functions, some of which have only recently been suggested and perhaps others which are as yet unrecognized. Among recognized unique features of albumin are: a) binding, and hence, inactivation of toxic products; b) regulation of the plasma and interstitial fluid concentrations of endogenous and exogenously administered substances and drugs; c) involvement in anticoagulation; d) maintenance of microvascular permeability to protein; and e) scavenging of free radicals and prevention of lipid peroxidation. This latter property may prove to be critically important, particularly in inflammatory disease states in which free radicals are thought to be a major culprit in direct damage due to tissue oxidation, and indirect tissue damage due to inactivation of important antiproteinases such as α_1 -PI and AT-III.

20

The following is a more detailed summary of the many uses for ALB that have been reported in the literature:

25

A. Functions of ALB

30

- Contributes to colloid osmotic pressure and thus prevents water loss from circulation;

35

- Aids in transport, distribution, metabolism of fatty acids (primarily long chain), amino acids (Cys and Trp), steroids, glutathione, metals (Ca, Zn), bilirubin, lysolecithin, hematin, prostaglandins and pharmaceuticals to liver, intestine, kidney and brain presumably through specific albumin receptors that have been identified on the endothelium;

- Serves as a reservoir for fatty acids intra and extravascularly (60% of the ALB is found extravascularly);
5
- Modification of doxorubicin (DXR) by conjugating it to bovine serum albumin (BSA) improved chemotherapeutic efficiency of DXR presumably by decreasing efflux of BSA-DXR compared to DXR alone (in 10 animal models), suggesting a similar use with ALB;
- Inhibits Cu-stimulated lipid peroxidation and hemolysis of erythrocyte membranes (acts as antioxidant);
15
- Scavenges HOCl and peroxy radicals;
- Prevents peroxidation of fatty acids by binding to them;
20
- May exert a protective effect in body fluids that have little endogenous antioxidant protection (e.g., eye and cerebrospinal fluids);
25
- In urine, high levels of ALB are diagnostic for detection of early renal pathology in diabetics;
- Administered to combat shock and given to 30 neonates with respiratory distress syndrome;
- Administered as a vehicle for hematin to treat acute intermittent porphyria;
35
- Used in tissue culture in place of whole serum;

- Enhances effectiveness of superoxide dismutase (SOD) when coupled to SOD through enhanced serum half-life;

5

- In microsphere form, ALB is useful as a carrier of therapeutic agents;

10

- Inhibits hydrogen peroxide release and neutrophil spreading.

B. Publications relating to ALB

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II. Alpha Fetoprotein

10 Alpha-fetoprotein (AFP; molecular weight 70,000) is a major serum protein produced during development and is produced primarily by the fetal liver and yolk sac cells. Its synthesis decreases markedly after birth and only trace amounts are present in the 15 serum of adults. Increased adult serum levels are a sign of hepatoma or yolk sac tumor, since these tumors produce AFP. The specific associations of AFP with fetal development as well as the above type of malignancies has attracted much interest and many 20 studies have been done on the structure of AFP and its gene, the regulation of gene expression, and biological functions.

25 Similar to ALB, AFP has been shown to bind various ligands such as unsaturated fatty acids, estrogens, bilirubin, copper and nickel ions, and others. AFP also has been claimed to regulate immune processes in a variety of systems from many different laboratories, although the results are controversial.

30 The following is a more detailed list of uses for AFP that are available in the literature:

A. Functions of AFP

- Binds unsaturated fatty acids, estrogens, 35 bilirubin, Cu, Ni;

- Elevated levels in amniotic fluid of pregnant women indicative of fetal malformations;
- High levels also found in hereditary 5 tyrosinemia and ataxia-telangiectasia (autosomal recessive disorder characterized by a defect in tissue differentiation of thymus and liver);
- Inhibits NK cell activity; 10
- Induces T suppressor cells;
- Inhibits mitogenic responses of 15 lymphocytes to PHA and ConA;
- Inhibits T cell proliferation to Ia determinants; 20
- Decreases macrophage phagocytosis and Ia expression;
- Inhibits FSH-mediated estradiol production by porcine granulosa cells; 25
- Enhances growth-factor mediated cell proliferation of porcine granulosa cells.

B. Publications relating to AFP

30 48. Suzuki, Y., et al., *J. Clin. Invest.* 90, 1530-1536 (1992).

49. Sakai, M., et al., *J. Biol. Chem.* 260 (8), 5055-5060 (1985).

50. *EPO Patent Application No. 0353814,*
February 7, 1990.

III. Vitamin-D Binding Protein

5

The group-specific component (Gc; VDB) is an α_2 -globulin of molecular weight 51,000. It is synthesized in the liver and is the major vitamin D-binding protein in plasma. VDB appears in human 10 populations as three common genetic phenotypes: Gc1, Gc2, and Gc2-1. VDB has also been reported to bind G-actin and to be spatially associated with IgG on lymphocyte membranes.

The following is a more detailed list of uses 15 for VDB that are available in the literature:

A. Functions of VDB

• Binds seco-steroid, vitamin D and the 20 derivatives 25-hydroxy vitamin D and 1,25 hydroxy vitamin D, possibly for transport in plasma;

• 1,25 vitamin D can differentiate monocytes and VDB prevents this;

25 • Binds actin (prevents assembly of actin polymers);

• Binds unsaturated fatty acids (e.g., 30 arachidonic acid);

• Binds C5a and C5a des Arg to act as a chemotactic factor for neutrophils;

35 • Acts as an activating factor for macrophages.

B. Publication relating to VDB

51. Watt, G. H., et al., *Circulatory Shock*
5 28, 279-291 (1989).

The protein of the present invention, AFM, bears a strong similarity in structure to ALB, AFP, and VDB, and is therefore expected to share the above 10 utilities and activities with the known proteins discussed above.

Summary of the Invention

15 In the course of experiments designed to purify a serum protein which could inhibit the binding of lipopolysaccharide (LPS)-coated erythrocytes to human macrophages, the inventors purified a novel human protein that co-purifies with apolipoprotein A1 (ApoA1).
20 The novel protein has an apparent molecular weight of 87,000 when run on SDS-PAGE and is designated as AFM. Herein, the inventors describe the cloning of the cDNA for AFM and demonstrate that AFM has a striking similarity, both structurally and functionally, to other 25 members of the ALB family. In addition, the inventors purified AFM from the serum-free conditioned medium of CHO D⁻ cells transfected with the cDNA for AFM, thus allowing the study of AFM in the absence of ApoA1.

Based on the above, the present invention 30 provides purified and isolated polynucleotides (e.g., DNA sequences and RNA transcripts thereof) encoding a novel human polypeptide, "AFM" as well as complexes of AFM with ApoA1 and/or lipids, and polypeptide variants (including fragments and analogs) thereof which display 35 one or more biological activities or properties specific to AFM.

Preferred DNA sequences of the invention include genomic and cDNA sequences as well as wholly or partially chemically and or enzymatically synthesized DNA sequences and biological replicas thereof. Also provided are autonomously replicating recombinant constructions such as plasmid and viral DNA vectors incorporating such sequences and especially vectors, wherein DNA encoding AFM or an AFM variant are operatively linked to an endogenous or exogenous expression control DNA sequence.

According to another aspect of the invention, host cells, especially unicellular host cells such as prokaryotic and eukaryotic cells, are stably transformed with DNA sequences of the invention in a manner allowing AFM and variants thereof to be expressed therein.

Host cells of the invention are useful in methods for the large scale production of AFM and AFM variants wherein the cells are grown in a suitable culture medium and the desired polypeptide products are isolated from the cells or from the growth medium.

Novel AFM and AFM variant products of the invention may be obtained as isolates from natural cell sources, but are preferably produced by recombinant procedures involving host cells of the invention. The products may be obtained in fully or partially glycosylated, partially or wholly deglycosylated, or non-glycosylated forms, depending on the host cell selected for recombinant production and/or post-isolation processing. The products may also be bound to other molecules, such as cellularly derived lipids and/or ApoA1.

Products of the invention include monomeric and multimeric polypeptides having the sequence of amino acid residues numbered -21 through 578 as set out in FIG. 1 herein. As explained in detail infra, this sequence includes a putative signal or leader sequence

which precedes the "mature" protein sequence and spans residue -21 (Met) through residue -1 (Thr) followed by the mature protein spanning residues 1 (Leu) to residue 578 (Asn). Based on amino acid composition, the 5 calculated molecular weight of the mature protein lacking glycosylation or other post-translational modification is approximately 66,576 daltons.

AFM variants of the invention may comprise fragments including one or more of the regions specified 10 herein and may also comprise polypeptide analogs wherein one or more of the specified amino acids is deleted or replaced: (1) without substantial loss, and preferably with enhancement, of one or more biological activities or immunological characteristics specific for AFM; or 15 (2) with specific modulation of a particular ligand/receptor binding function. Analog polypeptides including additional amino acid residues (e.g., lysine) that facilitate multimer formation are also contemplated.

20 Further comprehended by the present invention are antibodies (e.g., monoclonal and polyclonal antibodies, single chain antibodies, chimeric antibodies, CDR-grafted antibodies and the like) or other binding proteins which are specific for AFM or AFM 25 variants. Antibodies can be developed using isolated natural or recombinant AFM or AFM variants.

30 The antibodies are useful in complexes for immunization as well as for purifying polypeptides of the invention. The antibodies are also useful in modulating (i.e., blocking, inhibiting or stimulating) 35 ligand/receptor binding reactions involving AFM.

Anti-idiotypic antibodies specific for anti-AFM antibodies and uses of such anti-idiotypic antibodies in treatment are also contemplated. Assays 35 for the detection and quantification of AFM on cell surfaces and in fluids such as serum may involve a

single antibody or multiple antibodies in a "sandwich" assay format.

The uses of the DNA and amino acid sequences of the present invention are varied. For example, 5 knowledge of the sequence of a cDNA for AFM makes possible the isolation by DNA/DNA hybridization of genomic DNA sequences encoding AFM and specifying AFM expression control regulatory sequences such as promoters, operators and the like. DNA/DNA 10 hybridization procedures carried out with DNA sequences of the invention and under stringent conditions are likewise expected to allow the isolation of DNAs encoding allelic variants of AFM, other structurally related proteins sharing the biological and/or 15 immunological specificity of AFM, and proteins homologous to AFM from non-human species (especially from other mammals). DNAs of the invention are useful in DNA/RNA hybridization assays to detect the capacity of cells to synthesize AFM. A variety of specific uses 20 for AFM are disclosed herein below. These uses are primarily based on the known uses of the homologous albumin type polypeptides discussed above.

Also made available by the invention are anti-sense polynucleotides (e.g., DNA and RNA) relevant to 25 regulating expression of AFM by those cells which ordinarily express it. Furthermore, knowledge of the DNA and amino acid sequences of AFM make possible the generation by recombinant means of hybrid fusion proteins characterized by the presence of AFM protein 30 sequences and immunoglobulin heavy chain constant regions and/or hinge regions. See, Capon, et al., *Nature*, 337: 525-531 (1989); Ashkenazi, et al., *P.N.A.S.* (USA), 88: 10535-10539 (1991); and *PCT WO 89/02922*, published April 6, 1989.

BRIEF DESCRIPTION OF THE FIGURES

Numerous other aspects and advantages of the present invention will therefore be apparent upon 5 consideration of the following detailed description thereof, reference being made to the drawings wherein:

FIG. 1 shows the nucleotide and deduced amino acid sequence of AFM. The putative signal sequence is indicated in lower case letters. Asterisks indicate 10 putative sites for N-glycosylation. These are also represented as SEQ ID NO:1 AND SEQ ID NO:2.

FIG. 2A and 2B show a comparison of ALB family amino acid sequences. FIG. 2A shows the alignment of ALB family proteins. Sequences were aligned using the 15 Clustal method in the MegAlign program (DNASTAR). Identical amino acid residues are boxed. Consensus indicates residues identical in all 4 sequences. Majority indicates 2 or 3 residues identical in all 4 sequences. FIG. 2B shows percent similarity (right of 20 diagonal) and identity (left of diagonal) between ALB family members. Similarities were determined using the GCG GAP program. The sequences for the comparison proteins are also provided as follows: serum albumin, SEQ ID NO:3; alpha fetoprotein, SEQ ID NO:4; and vitamin 25 D binding protein, SEQ ID NO:5.

FIG. 3 shows the conserved Cys pattern in ALB family proteins. The mature form of ALB family proteins are depicted with thin vertical bars representing single Cys residues and thick vertical lines representing -Cys- 30 Cys- sequences.

FIG. 4 shows the putative disulfide bonding pattern for AFM. The organization of domains and double loops are drawn as originally proposed for ALB.

FIG. 5 shows the expression of rAFM in stably transfected CHO D⁻ cells. Samples were applied to SDS-PAGE under reducing conditions followed by 5 electrophoretic transfer to nitrocellulose. After blocking with skim milk, the membrane was probed with the AM339 antibody followed by incubation with donkey rabbit anti-Ig. Immunoreactive proteins were visualized by chemiluminescence. Lane 1, 80 ml conditioned medium 10 (CM) from CHO D⁻ cells transfected with AFM cDNA; lane 2, 80 ml CM from nontransfected CHO D⁻ cells; lane 3, 100 ng AFM purified from human plasma. Size markers (in kDa) are indicated on the left.

FIG. 6 shows purification of rAFM. Samples 15 purified from CHO D⁻ cells transfected with AFM cDNA were adjusted with an equal volume of 2X sample buffer (125 mM Tris-HCl, pH 6.8, 4% SDS, 0.005% bromophenol blue, 10% glycerol) and analyzed by SDS-PAGE using 4-20% polyacrylamide gradient gels (Novex) under reducing 20 conditions. The gel was stained with Coomassie Brilliant Blue. Lanes 1 and 6, 10 mg Mark-12 molecular weight markers (Novex, Inc.); lane 2, 50 mg total protein after addition of ammonium sulfate to concentrated CM 25 (supernatant was dialyzed against PBS and subsequently loaded onto the gel); lane 3, 25 mg Phenyl Sepharose water eluate; lane 4, 10 mg Q Sepharose-purified rAFM; lane 5, 1 mg of Superdex 200-purified rAFM.

30 DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

Definition of AFM

AFM is defined as a polypeptide having a qualitative biological activity or property in common 35 with AFM of FIG. 1.

Included within the scope of AFM as that term is used herein is the AFM having the amino acid sequence of AFM as set forth in FIG. 1, SEQ ID NO:2; glycosylated, deglycosylated or unglycosylated 5 derivatives of AFM; and lipidated or delipidated forms of AFM.

Also included within the scope of AFM are AFMs from any species, including without limitation: human, mouse, rat, pig, rabbit, monkey, dog, etc. Especially 10 preferred is the human form of AFM.

Variants of AFM

Variants of AFM include homologous amino acid sequence variants of the sequence of FIG. 1, and 15 homologous in-vitro-generated variants and derivatives of AFM, which are capable of exhibiting a biological activity or property in common with AFM of FIG. 1.

"Homologous" is used herein to refer to the residues in a candidate sequence that are identical with 20 the residues in the sequence of AFM in FIG. 1 after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent homology.

A biological activity or property of AFM is defined as either 1) immunological cross-reactivity with 25 at least one epitope of AFM, or 2) the possession of at least one regulatory or effector function qualitatively in common with AFM. Examples of those activities may be found in the section herein describing uses of AFM.

"Immunologically cross-reactive" as used 30 herein means that the candidate polypeptide is capable of competitively inhibiting the qualitative biological activity of AFM or an AFM variant having this activity with polyclonal antisera raised against the known active analog. Such antisera are prepared in conventional 35 fashion by injecting animals such as goats or rabbits, for example, subcutaneously with the known active analog

in complete Freund's adjuvant, followed by booster intraperitoneal or subcutaneous injection in incomplete Freund's adjuvant. An example of production of polyclonal antisera production is presented in the 5 examples section below.

Amino acid sequence variants of AFM are prepared with various objectives in mind, including increasing the affinity of AFM for its binding partner, facilitating the stability, purification and preparation 10 of AFM, and the like.

Amino acid sequence variants of AFM fall into one or more of three classes: insertional, substitutional, or deletional variants. These variants ordinarily are prepared by site specific mutagenesis of 15 nucleotides in the DNA encoding AFM, by which DNA encoding the variant is obtained, and thereafter expressing the DNA in recombinant cell culture. However, variant AFM fragments having up to about 100-150 amino acid residues are prepared conveniently by in 20 vitro synthesis.

The amino acid sequence variants of AFM are predetermined variants not found in nature or naturally occurring alleles. AFM variants typically exhibit the same qualitative biological activity as the naturally 25 occurring AFM molecule. However, AFM variants and derivatives that are not capable of binding to their ligands are useful nonetheless (a) as a reagent in diagnostic assays for AFM or antibodies to AFM, (b) when insolubilized in accordance with known methods, as 30 agents for purifying anti-AFM antibodies from antisera or hybridoma culture supernatants, and (c) as immunogens for raising antibodies to AFM or as immunoassay kit components (labeled, as a competitive reagent for the native AFM or unlabeled as a standard for AFM assay) so 35 long as at least one AFM epitope remains active.

While the site for introducing an amino acid sequence variation is predetermined, the mutation per se need not be predetermined. For example, in order to optimize the performance of a mutation at a given site, 5 random or saturation mutagenesis (where all 20 possible residues are inserted) is conducted at the target codon and the expressed AFM variant is screened for the optimal combination of desired activities. Such screening is within the ordinary skill in the art.

10 Amino acid insertions usually will generally be on the order of about from 1 to 10 amino acid residues; substitutions are typically introduced for single residues; and deletions will generally range about from 1 to 30 residues. Deletions or insertions 15 preferably are made in adjacent pairs, i.e. a deletion of 2 residues or insertion of 2 residues. It will be amply apparent from the following discussion that substitutions, deletions, insertions or any combination thereof are introduced or combined to arrive at a final 20 construct.

Insertional amino acid sequence variants of AFM are those in which one or more amino acid residues extraneous to AFM are introduced into a predetermined site in the target AFM and which displace the 25 preexisting residues. Commonly, insertional variants are fusions of heterologous proteins or polypeptides to the amino or carboxyl terminus of AFM. Such variants are referred to as fusions of AFM and a polypeptide containing a sequence which is other than that which is 30 normally found in AFM at the inserted position. Several groups of fusions are contemplated herein.

Immunologically active AFM derivatives and fusions comprise AFM and a polypeptide containing a non-AFM epitope, and are within the scope of this invention. 35 The non-AFM epitope is any immunologically competent polypeptide, i.e., any polypeptide which is capable of

eliciting an immune response in the animal to which the fusion is to be administered or which is capable of being bound by an antibody raised against the non-AFM polypeptide. Typical non-AFM epitopes will be those 5 which are borne by allergens, autoimmune epitopes, or other potent immunogens or antigens recognized by pre-existing antibodies in the fusion recipient, including bacterial polypeptides such as trpLE, beta-galactosidase, viral polypeptides such as herpes gD 10 protein, and the like.

15 Immunogenic fusions are produced by cross-linking in vitro or by recombinant cell culture transformed with DNA encoding an immunogenic polypeptide. It is preferable that the immunogenic fusion be one in which the immunogenic sequence is joined to or inserted into AFM or fragment thereof by a peptide bond(s). These products therefore consist of a linear polypeptide chain containing AFM epitope and at least one epitope foreign to AFM. It will be understood 20 that it is within the scope of this invention to introduce the epitopes anywhere within the AFM molecule or fragment thereof.

25 Such fusions are conveniently made in recombinant host cells or by the use of bifunctional cross-linking agents. The use of a cross-linking agent to fuse AFM to the immunogenic polypeptide is not as desirable as a linear fusion because the cross-linked products are not as easily synthesized in structurally homogeneous form.

30 These immunogenic insertions are particularly useful when formulated into a pharmacologically acceptable carrier and administered to a subject in order to raise antibodies against AFM, which antibodies in turn are useful in diagnostics or in purification of 35 AFM by immunoaffinity techniques known per se. In diagnostic applications, the antibodies will typically

be bound to or associated with a detectable group, examples of which are well known to those skilled in the art. Immunoaffinity techniques could be used, for example, to purify AFM.

5 Other fusions, which may or may not also be immunologically active, include fusions of the mature AFM sequence with a signal sequence heterologous to AFM, and fusions of AFM to polypeptides having enhanced plasma half life (ordinarily > about 20 hours) such as
10 immunoglobulin chains or fragments thereof.

Signal sequence fusions are employed in order to more expeditiously direct the secretion of AFM. The heterologous signal replaces the native AFM signal, and when the resulting fusion is recognized, i.e. processed
15 and cleaved by the host cell, AFM is secreted. Signals are selected based on the intended host cell, and may include bacterial yeast, mammalian and viral sequences. The native AFM signal or the herpes gD glycoprotein signal is suitable for use in mammalian expression
20 systems.

Substantial variants are those in which at least one residue in the FIG. 2 sequence has been removed and a different residue inserted in its place. Such substitutions generally are made in accordance with
25 the following Table 1 when it is desired to finely modulate the characteristics of AFM.

TABLE 1

Original Residue	Exemplary Substitutions
Ala	Ser
Arg	Lys
Asn	Gln; His
Asp	Glu
Cys	Ser; Ala
Gln	Asn
Glu	Asp
Gly	Pro
His	Asn; Gln
Ile	Leu; Val
Leu	Ile; Val
Lys	Arg; Gln; Glu
Met	Leu; Ile
Phe	Met; Leu; Tyr
Ser	Thr
Thr	Ser
Trp	Tyr
Tyr	Trp; Phe
Val	Ile; Leu

Substantial changes in function or immunological identity are made by selecting substitutions that are less conservative than those in 5 Table 1, i.e., selecting residues that differ more significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example as a sheet or helical conformation, (b) the charge or hydrophobicity of the 10 molecule at the target site or (c) the bulk of the side chain. The substitutions which in general are expected to produce the greatest changes in AFM properties will

be those in which (a) a hydrophilic residue, e.g. seryl or threonyl, is substituted for (or by) a hydrophobic residue, e.g. leucyl, isoleucyl, phenylalanyl, valyl or alanyl; (b) a cysteine or proline is substituted for (or by) any other residue; (c) a residue having an electropositive side chain, e.g., lysyl, arginyl, or histidyl, is substituted for (or by) an electronegative residue, e.g., glutamyl or aspartyl; or (d) a residue having a bulky side chain, e.g., phenylalanine, is substituted for (or by) one not having a side chain, e.g. glycine.

Some deletions, insertions, and substitutions will not produce radical changes in the characteristics of AFM molecule. However, when it is difficult to predict the exact effect of the substitution, deletion, or insertion in advance of doing so, for example when modifying the AFM extracellular domain or an immune epitope, one skilled in the art will appreciate that the effect will be evaluated by routine screening assays.

Another class of AFM variants are deletional variants. Deletions are characterized by the removal of one or more amino acid residues from AFM sequence. Deletions from the AFM C-terminal or the N-terminal, which preserve the biological activity or immune cross-reactivity of AFM are suitable.

Deletions of cysteine or other labile residues also may be desirable, for example in increasing the oxidative stability of AFM. Deletion or substitutions of potential proteolysis sites, e.g. Arg-Arg, is accomplished by deleting one of the basic residues or substituting one by glutamyl or histidyl residues.

Preferably, the variants represent conservative substitutions. It will be understood that some variants may exhibit reduced or absent biological activity. These variants nonetheless are useful as

standards in immunoassays for AFM so long as they retain at least one immune epitope of AFM.

Glycosylation variants are included within the scope of AFM. They include variants completely lacking in glycosylation (nonglycosylated) and variants having at least one less glycosylated site than the native form (deglycosylated) as well as variants in which the glycosylation has been changed. Additionally, unglycosylated AFM which has the amino acid sequence of the native AFM is produced in recombinant prokaryotic cell culture because prokaryotes are incapable of introducing glycosylation into polypeptides.

Glycosylation variants are produced by selecting appropriate host cells or by in vitro methods. Yeast, for example, introduce glycosylation which varies significantly from that of mammalian systems. Similarly, mammalian cells having a different species (e.g. hamster, murine, insect, porcine, bovine or ovine) or tissue origin (e.g. lung, lymphoid, mesenchymal or epidermal) than the source of AFM are routinely screened for the ability to introduce variant glycosylation as characterized for example by elevated levels of mannose or variant ratios of mannose, fucose, sialic acid, and other sugars typically found in mammalian glycoproteins. In vitro processing of AFM typically is accomplished by enzymatic hydrolysis, e.g. neuraminidase digestion.

AFM isolated from natural sources or produced recombinantly will generally contain bound lipids. The nature of the bound lipids is expected to depend on the source of the AFM. Delipidated versions of AFM may be prepared by standard delipidation methods known in the art, especially the art relating to ALB where delipidation is a common procedure. One preferred method involves extracting an aqueous solution of AFM with a solvent capable of dissolving lipids, such as 1-

butanol or diisopropyl ether. Example 8 presents a specific exemplary method for delipidation.

Certain post-translational derivatizations are the result of the action of recombinant host cells on 5 the expressed polypeptide. Glutaminyl and asparaginyl residues are frequently post-translationally deamidated to the corresponding glutamyl and aspartyl residues. Alternatively, these residues are deamidated under mildly acidic conditions. Either form of these residues 10 falls within the scope of this invention.

Other post-translational modifications include hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the α -amino groups of lysine, arginine, 15 and histidine side chains (T. E. Creighton, *Proteins: Structure and Molecular Properties*, (W. H. Freeman & Co.), San Francisco: 79-86 (1983), acetylation of the N-terminal amine and, in some instances, amidation of the C-terminal carboxyl.

20

Oligonucleotides relating to AFM

DNA encoding AFM is synthesized by in vitro methods or is obtained readily from human liver cDNA libraries. The means for synthetic creation of the DNA 25 encoding AFM, either by hand or with an automated apparatus, are generally known to one of ordinary skill in the art, particularly in light of the teachings contained herein. As examples of the state of the art relating to polynucleotide synthesis, one is directed to 30 Maniatis et al., *Molecular Cloning-A Laboratory Manual*, Cold Spring Harbor Laboratory (1984), and Horvath et al., *An Automated DNA Synthesizer Employing Deoxynucleoside 3'-Phosphoramidites, Methods in Enzymology* 154: 313-326 (1987).

35

Alternatively, to obtain DNA encoding AFM from sources other than murine or human, since the entire DNA

sequence for the preferred embodiment of AFM is given, one needs only to conduct hybridization screening with labelled DNA encoding AFM or fragments thereof (usually, greater than about 20, and ordinarily about 50bp) in 5 order to detect clones which contain homologous sequences in the cDNA libraries derived from the liver of the particular animal, followed by analyzing the clones by restriction enzyme analysis and nucleic acid sequencing to identify full-length clones. If full 10 length clones are not present in the library, then appropriate fragments are recovered from the various clones and ligated at restriction sites common to the fragments to assemble a full-length clone. DNA encoding AFM from other animal species is obtained by probing 15 libraries from such species with the human sequences, or by synthesizing the genes in vitro.

Included within the scope hereof are nucleic acid sequences that hybridize under stringent conditions to a fragment of the DNA sequence in FIG. 1, which 20 fragment is greater than about 10 bp, preferably 20-50 bp, and even greater than 100 bp. Also included within the scope hereof are nucleic acid sequences that hybridize under stringent conditions to a fragment of AFM. "Stringent" is used to refer to conditions that 25 are commonly understood in the art as stringent. An exemplary set of conditions include a temperature of 60 - 70°C, (preferably about 65°C) and a salt concentration of 0.70M to 0.80M (preferably about 0.75M).

Included also within the scope hereof are 30 nucleic acid probes which are capable of hybridizing under stringent conditions to the cDNA of AFM or to the genomic gene for AFM (including introns and 5' or 3' flanking regions extending to the adjacent genes or about 5,000 bp, whichever is greater).

Recombinant Expression of AFM

In general, prokaryotes are used for cloning of DNA sequences in constructing the vectors useful in the invention. For example, *E. coli* K12 strain 294 (ATCC No. 31446) is particularly useful. Other microbial strains which may be used include *E. coli* B and *E. coli* X1776 (ATCC No. 31537). Alternatively, *in vitro* methods of cloning, e.g. polymerase chain reaction, are suitable.

AFMs of this invention are expressed directly in recombinant cell culture as an N-terminal methionyl analog, or as a fusion with a polypeptide heterologous to AFM, preferably a signal sequence or other polypeptide having a specific cleavage site at the N-terminus of AFM. For host prokaryotes that do not process AFM signal, the signal is substituted by a prokaryotic signal selected for example from the group of the alkaline phosphatase, penicillinase, or heat stable enterotoxin II leaders. For yeast secretion the human AFM signal may be substituted by the yeast invertase, alpha factor or acid phosphatase leaders. In mammalian cell expression the native signal is satisfactory for mammalian AFM, although other mammalian secretory protein signals are suitable, as are viral secretory leaders, for example the herpes simplex gD signal.

AFM may be expressed in any host cell, but preferably are synthesized in mammalian hosts. However, host cells from prokaryotes, fungi, yeast, insects and the like are also used for expression. Exemplary prokaryotes are the strains suitable for cloning as well as *E. coli* W3110 (F-, l-, prototrophic, ATCC No. 7325), other enterobacteriaceae such as *Serratia marescans*, bacilli and various pseudomonads. Preferably the host cell should secrete minimal amounts of proteolytic enzymes.

Expression hosts typically are transformed with DNA encoding AFM which has been ligated into an expression vector. Such vectors ordinarily carry a replication site (although this is not necessary where chromosomal integration will occur). Expression vectors also include marker sequences which are capable of providing phenotypic selection in transformed cells, as will be discussed further below. Expression vectors also optimally will contain sequences which are useful for the control of transcription and translation, e.g., promoters and Shine-Dalgarno sequences (for prokaryotes) or promoters and enhancers (for mammalian cells). The promoters may be, but need not be, inducible.

Promoters suitable for use with prokaryotic hosts illustratively include the β -lactamase and lactose promoter systems (Chang et al., *Nature*, 275, 615 (1978); and Goeddel et al., *Nature* 281, 544 (1979)), alkaline phosphatase, the tryptophan (trp) promoter system (Goeddel, *Nucleic Acids Res.* 8: 4057 (1980) and EPO *Appln. Publ.* No. 36,776) and hybrid promoters such as the tac promoter (H. de Boer et al., *Proc. Natl. Acad. Sci. USA* 80, 21-25 (1983)). However, other functional bacterial promoters are suitable. Their nucleotide sequences are generally known, thereby enabling a skilled worker operably to ligate them to DNA encoding AFM (Siebenlist et al., *Cell* 20, 269 (1980)) using linkers or adapters to supply any required restriction sites. Promoters for use in bacterial systems also will contain a Shine-Dalgarno (S.D.) sequence operably linked to the DNA encoding AFM.

In addition to prokaryotes, eukaryotic microbes such as yeast or filamentous fungi are satisfactory. *Saccharomyces cerevisiae* is the most commonly used eukaryotic microorganism, although a number of other strains are commonly available.

Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., *J. Biol. Chem.* **255**, 2073 (1980)) or other glycolytic enzymes (Hess et al., 5 *J. Adv. Enzyme Reg.* **7**: 149 (1968); and Holland, *Biochemistry* **17**, 4900 (1978)), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate 10 kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the 15 promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde 3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. 20 Suitable vectors and promoters for use in yeast expression are further described in R. Hitzeman et al., *European Patent Publication No. 73,657A*.

Expression control sequences are known for eukaryotes. Virtually all eukaryotic genes have an AT-rich region located approximately 25 to 30 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide. At the 3' end of most 25 eukaryotic genes is an AATAAA sequence which may be the signal for addition of the poly A tail to the 3' end of the coding sequence. All of these sequences are 30 inserted into mammalian expression vectors.

Suitable promoters for controlling 35 transcription from vectors in mammalian host cells are readily obtained from various sources, for example, the

genomes of viruses such as polyoma virus. SV40, adenovirus, MMV (steroid inducible), retroviruses (e.g. the LTR of HIV), hepatitis-B virus and most preferably cytomegalovirus, or from heterologous mammalian promoters, e.g. the beta actin promoter. The early and late promoters of SV40 are conveniently obtained as an SV40 restriction fragment which also contains the SV40 viral origin of replication. Fiers et al., *Nature* 273, 113 (1978). The immediate early promoter of the human cytomegalovirus is conveniently obtained as a HindIII E restriction fragment. Greenaway, P. J. et al., *Gene* 18, 355-360 (1982).

Transcription of a DNA encoding AFM by higher eukaryotes is increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10-300 bp, that act on a promoter to increase its transcription. Enhancers are relatively orientation and position independent having been found 5' (Laimins, L. et al., *PNAS* 78, 993 (1981)) and 3' (Lusky, M. L., et al., *Mol. Cell Bio.* 3, 1108 (1983) to the transcription unit, within an intron (Banerji, J. L. et al., *Cell* 33: 729 (1983)) as well as within the coding sequence itself (Osborne, T. F., et al., *Mol. Cell Bio.* 4, 1293 (1984)). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of

transcription which may affect mRNA expression. These regions are transcribed as polyadenylated segments in the untranslated portion of the mRNA encoding AFM. The 3' untranslated regions also include transcription 5 termination sites.

Expression vectors may contain a selection gene, also termed a selectable marker. Examples of suitable selectable markers for mammalian cells are dihydrofolate reductase (DHFR), thymidine kinase (TK) or 10 neomycin.

Suitable eukaryotic host cells for expressing AFM include monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, 15 Graham, F. L. et al., *J. Gen Virol.* 36: 59 (1977)); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary-cells-DHFR (CHO, Urlaub and Chasin, *PNAS (USA)* 77, 4216, (1980)); mouse sertoli cells (TM4, Mather, J. P., *Biol. Reprod.* 23, 243-251 (1980)); monkey kidney cells 20 (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver 25 cells (Hep G2, HB 8065); mouse mammary tumor (MMT 060562, ATCC CCL51); and, TRI cells (Mather, J. P. et al., *Annals N.Y. Acad. Sci.* 383, 44-68 (1982)).

Construction of suitable vectors containing the desired coding and control sequences employ standard 30 ligation techniques. Isolated plasmids or DNA fragments are cleaved, tailored, and religated in the form desired to form the plasmids required.

Host cells are transformed with the expression vectors of this invention and cultured in conventional 35 nutrient media modified as appropriate for inducing promoters, selecting transformants or amplifying AFM

gene. The culture conditions, such as temperature, pH and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

5 The host cells referred to in this disclosure encompass cells in in vitro culture as well as cells which are within a host animal.

"Transformation" means introducing DNA into an organism so that the DNA is replicable, either as an 10 extrachromosomal element or by chromosomal integration. The method used herein for transformation of the host cells may be, for example, the method of Graham, F. and van der Eb, A., *Virology* 52, 456-457 (1973). However, other methods for introducing DNA into cells such as by 15 nuclear injection or by protoplast fusion may also be used. If prokaryotic cells or cells which contain substantial cell wall constructions are used, the preferred method of transfection is calcium treatment using calcium chloride as described by Cohen, F. N. 20 et al., *Proc. Natl. Acad. Sci. (USA)* 69, 2110 (1972).

"Transfection" refers to the introduction of DNA into a host cell whether or not any coding sequences are ultimately expressed. Numerous methods of transfection are known to the ordinarily skilled artisan, for example, CaPO₄ and electroporation. 25 Transformation of the host cell is indicative of successful transfection.

Recovery and Purification of AFM

30 AFM is recovered and purified from recombinant cell cultures by known methods, including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, immunoaffinity chromatography, 35 hydroxyapatite chromatography, lectin chromatography, hydrophobic interaction chromatography, and gel

filtration chromatography. Other known purification methods within the scope of this invention utilize, for example, immobilized carbohydrates, epidermal growth factor, or complement domains. Moreover, reverse-phase 5 HPLC and chromatography using anti-AFM antibodies are useful for the purification of AFM. AFM may preferably be purified in the presence of a protease inhibitor such as PMSF. A specific preferred method of purifying AFM is found in Example 1.

10

Uses of AFM

As mentioned above, in view of the homology between AFM and each of ALB, AFP and VDB, it is expected that AFM and variants thereof will have identical or 15 similar biological activities and utilities. In general, members of the albumin family show a high propensity to bind and transport a wide variety of substances through the body including fatty acids, hormones, enzymes, dyes, trace metals and drugs. It is expected that a 20 preparation of AFM which has its endogenous lipids removed could be used to reduce high concentrations of free fatty acids (hyperlipidemia) found in disease states such as acute pancreatitis, ARDS, sepsis and atherosclerosis.

25 Similar to proposed functions for ALB, AFM can also potentially be used as an antioxidant. AFM is expected to act as an antioxidant by sequestering metal ions and preventing those ions from accelerating free-radical reactions (ie. decomposition of lipid peroxides 30 to peroxy and alkoxy radicals; formation of hydroxyl radical from hydrogen peroxide) or AFM may act to directly inactivate hydrogen peroxide, hydroxyl radicals and hypochlorous acid. The detoxification of oxygen metabolites by AFM can limit the detrimental effects 35 that unbound oxygen metabolites have in inactivating beneficial anti-proteases (α_1 -antiproteinase and α_1 -

antitrypsin) and in damaging DNA, proteins and lipids. Thus, AFM may be used to ameliorate ischaemia-reperfusion injury, rheumatoid arthritis, ARDS, cardiopulmonary bypass, sepsis and any other diseases or 5 tissue damage caused by an excess production of oxygen metabolites from leukocytes and/or arachidonic acid metabolism.

Another use of AFM is based on the finding that albumin family proteins bind and detoxify a wide 10 variety of toxic products including man-made drugs. Thus, AFM can ameliorate the effects of toxic plasma substances released as a result of inflammation and can be conjugated to toxic pharmaceuticals so as to minimize detrimental activities of the drug.

15 AFM may also have anticoagulant properties as has been reported for ALB. AFM may inhibit platelet aggregation and thus could be a useful additive to resuscitation fluids in disease states characterized by increased platelet aggregation such as sepsis, 20 hemorrhagic shock and burn injury.

Serum AFM levels may increase or decrease due to a particular pathological condition as has been observed with AFP. Thus, calculation of serum AFM levels using antibodies described in Example 11 will be useful 25 in diagnosing specific human diseases.

Administration of AFM

For administration in vivo, AFM is placed into sterile, isotonic formulations, and administered by 30 standard means well known in the field. The formulation of AFM is preferably liquid, and is ordinarily a physiologic salt solution containing 0.5-10 mM calcium, non-phosphate buffer at pH 6.8-7.6, or may be lyophilized powder.

35 It is envisioned that intravenous delivery, or delivery through catheter or other surgical tubing will

be the primary route for therapeutic administration. Alternative routes include tablets and the like, commercially available nebulizers for liquid formulations, and inhalation of lyophilized or 5 aerosolized receptors. Liquid formulations may be utilized after reconstitution from powder formulations.

AFM may also be administered via microspheres, liposomes, other microparticulate delivery systems or sustained release formulations placed in certain tissues 10 including blood.

The dose of AFM administered will be dependent upon the properties of AFM employed, e.g. its activity and biological half-life, the concentration of AFM in the formulation, the administration route for AFM, the 15 site and rate of dosage, the clinical tolerance of the patient involved, the pathological condition afflicting the patient and the like, as is well within the skill of the physician.

AFM may also be administered along with other 20 pharmacologic agents such as antibiotics, anti-inflammatory agents, and anti-tumor agents. It may also be useful to administer AFM along with other therapeutic proteins such as gamma-interferon and other immunomodulators.

25

Ligands of AFM

It is also possible that the AFM of the present invention will have an additional natural ligand or ligands (other than the lipids discussed herein) and 30 that such ligands will be capable of modulating the biological activities of AFM in vivo. One of ordinary skill may be able to utilize the teachings of the present invention to screen for ligands of the AFMs disclosed herein and then isolate and purify them, e.g., 35 by immunochromatography using AFM or a variant of AFM bound to a solid support.

Antibodies to AFM

Also included within the scope of the invention are antibodies (e.g., monoclonal and 5 polyclonal antibodies, single chain antibodies, chimeric antibodies, CDR-grafted antibodies and the like) or other binding proteins which are specific for AFM or its variants. Antibodies can be developed using natural or recombinant AFM or AFM variants or cells expressing such 10 molecules on their surfaces. Active fragments of such antibodies are also contemplated.

The antibodies are useful for purifying the polypeptides described herein, or identifying and purifying cells producing the polypeptides on their 15 surfaces. The antibodies could also be used to modulate (e.g., block, inhibit or stimulate) ligand binding to AFM. Anti-idiotypic antibodies are also contemplated. Assays for detection and quantitation of AFM on cell surfaces and in fluids (such as serum, blood, 20 cerebrospinal fluid, urine, semen, milk, and tears) may involve a single antibody or a "sandwich" assay format.

* * *

25 It is understood that the application of the teachings of the present invention to a specific problem or situation will be within the capabilities of one having ordinary skill in the art in light of the teachings contained herein. Examples of the products of 30 the present invention and representative processes for their isolation, use, and manufacture appear below. All literature and patent citations herein are expressly incorporated by reference.

EXAMPLES

EXAMPLE 1: Lipopolysaccharide (LPS) Neutralization Assay

5 A. Formation of ELPS.

Sheep erythrocytes are first coated with *S. minnesota* Re595 LPS (List Biological) as previously described (J. Ex. Med. 170:1231). The concentration of LPS used to coat erythrocytes is $10 \text{ mg}/8 \times 10^7$ erythrocytes. After coating with LPS, erythrocytes are resuspended in 5 mM veronal buffer, pH 7.5, containing 150 mM NaCl, 0.1% gelatin and 1 mM EDTA (EDTAGVB²⁻) to a concentration of 1×10^8 cells/ml.

15 B. Opsonization of ELPS.

ELPS are opsonized using normal human plasma (NHP) as a source of opsonizing proteins (septin) (J. Ex. Med. 176:719) or LBP (J. Ex. Med. 170:1231). NHP is diluted 1:100 in phosphate buffered saline without divalent cations containing 1 mM EDTA (PDEDTA) and then mixed 1:1 with ELPS (1×10^8 cells/ml). The mixture is then incubated 10 minutes at 37°C, spun down for 1 minute at 800x g in a swinging bucket centrifuge. Cells are washed 2x with EDTAGVE²⁻ and resuspended to a final concentration of 1×10^8 cells/ml in EDTAGVB²⁻. Erythrocytes treated as above are denoted E-septins.

C. Assay of Samples for LPS Neutralizing Activity.

The following step measures the ability of a given sample to reduce the amount of binding of LPS coated erythrocytes to macrophages in a CD-14 dependent manner.

Dilutions of samples to be tested are made in PDEDTA. E-septins are incubated 1:1 with sample dilutions at 37°C for 40 minutes. One sample is included in which E-septin is incubated with PDEDTA alone as a control. Following incubation, E-septins are

spun down in a swinging bucket centrifuge at 800x g for 1 minute and resuspended in EDTAGVB²⁻ to a concentration of 1 x 10⁸ cells/ml. E-septins treated in this manner are denoted E-septin¹. Five ml of the resuspended E-septins¹ are added to a macrophage monolayer (see below) in a 60 well terasaki plate (NUNC, Inc.). The plate is incubated 20 minutes at room temperature followed by inversion and incubation another 12 minutes at room temperature to separate unbound E-septins¹ from the macrophages. The plate is then dipped in a beaker of cold PBS with no divalent cations containing 0.001% azide to wash off unbound erythrocytes. Binding of E-septins¹ is evaluated by phase contrast microscopy. Binding is expressed as the attachment index which denotes the number of erythrocytes bound per 100 macrophages. LPS neutralizing activity of the samples is evaluated by comparing the binding of E-septins¹ incubated with dilutions of the sample in PDEDTA to E-septin¹ incubated with PDEDTA alone. Activity is measured in units/ml. This represents the dilution at which the sample reduces E-septin¹ binding by 50% in the standard assay.

D. Macrophage Monolayer.

Monocyte derived macrophages are obtained by culturing human monocytes in teflon beakers as described (J. Ex. Med. 156:1149). On the day of the assay, macrophages are removed from the beaker, washed with PBS containing divalent cations (PBS²⁺) and resuspended in PBS²⁺ containing 0.5 mg/ml human serum albumin (Armour Pharm.), 0.3 U/ml aprotinin (Sigma) and 3 mM glucose (HAP buffer) to 1 x 10⁶ cells/ml. Thirty minutes prior to addition of E-septins¹ to the terasaki plate, macrophage monolayers are formed by adding 5 ml of cells and 5 ml of HAP buffer to each well. Immediately prior

to addition of E-septins¹, the plate is flooded with PDEDFA and the wells are lightly aspirated.

EXAMPLE 2: Purification of AFM

5 All steps, except where noted were done at controlled room temperature. All buffers were made with pyrogen free water and sterile filtered prior to use. One unit of fresh frozen plasma (FFP) was thawed by immersion of the packet in a room-temperature water
10 bath. The thawed plasma was first transferred to a graduated cylinder, the volume noted, and then poured into a beaker. The FFP was then stirred magnetically while the beaker was immersed in a ice-water bath. To the FFP, sufficient 3.9M ammonium sulfate (4°C) was
15 slowly added to achieve a final ammonium sulfate concentration of 1.6M. This mixture was allowed to stir at 4°C for an additional 4 hours. The mixture was then centrifuged at 10,000 x g for 60 minutes. The supernatant was recovered and allowed to come to room
20 temperature by immersion in a room temperature water bath.

A portion (250 ml) of the ammonium sulfate supernatant was loaded onto a 2.6 x 10cm Phenyl Sepharose HP column previously equilibrated with 50mM sodium phosphate, 1.6M ammonium sulfate; pH 7.5.
25 Loading, wash and elution steps were done at a linear flow rate of 40cm/hr. Following sample loading the column was washed in turn with 50mM sodium phosphate, 1.6M ammonium sulfate; pH 7.5 and 50mM sodium phosphate;
30 pH 7.5. Elution of activity (see Example 1) was accomplished by the introduction of water. The pH of the water eluate was adjusted to 8.2 with 0.1M Tris base and loaded, at a flow rate of 150cm/hr, onto a 2.6 x 10cm Q Sepharose HP column previously equilibrated with
35 20mM Tris-HCl; pH 8.2. Following sample loading the column was thoroughly washed with first 20mM Tris-HCl;

pH 8.2 and then 0.1M sodium chloride, 20mM Tris-HCl; pH 8.2. The column was then resolved with a 0.33mM/ml linear gradient of sodium chloride ranging from 0.1 to 0.3M, followed by a 7mM/ml linear gradient of sodium chloride ranging from 0.3M to 1.0M in 20mM Tris-HCl; pH 8.2. Active fractions were pooled (30 ml).

One third of the pooled activity from above was loaded onto a Superdex 200 prep column (2.6 x 60cm) previously equilibrated with phosphate buffered saline; pH 7.4. The column was loaded and resolved at a flow rate of 1ml/min. The fractions were analyzed for activity and by reducing SDS-PAGE. A pool (8ml) of the appropriate fractions showed two major protein bands by reducing SDS PAGE at a molecular weight of 87 kd and 28 kd. Two milliliters of the above pool was submitted for sequence analysis. The 28 kd protein had the same N-terminus as ApoA1 and the 87 kd protein had a novel N-terminus (see below). The remaining 6ml of the Superdex 200 prep pool was loaded onto a Superdex 200 prep column (2.6 x 60cm) previously equilibrated with 1% sodium deoxycholate, 0.15M sodium chloride, 50mM Tris-HCl; pH 8.5. The column was loaded and resolved at a flow rate of 1ml/min. The fractions were analyzed by SDS-PAGE. Those fractions corresponding to either the 87 kd or the 28 kd proteins were separately pooled (16ml each).

Four milliliters of each pool was diafiltered against 50mM Tris-HCl; pH 8.5 to remove excess sodium deoxycholate and concentrated to approximately 0.1mg/ml.

-34-

EXAMPLE 3: Amino Acid Sequence Analysis of N-terminus
and Proteolytic Fragments of Purified AFM

To obtain the N-terminal sequence, 10 ug of purified AFM was loaded on the reverse phase portion of a precycled biphasic sequencing column. The column was loaded on a Hewlett Packard G1004A protein sequencer with on-line phenylthiohydantoin amino acid analysis performed with a Hewlett Packard 1090 high performance liquid chromatography (HPLC). The N-terminal amino acid sequence obtained from the protein was:

LPTQPRDIENFXSTQKFIEDNIEYITIIAFAQYVQ (SEQ ID NO: 6),

where "X" represents an unassignable amino acid.

To obtain sequences from AFM tryptic peptide fragments, 100 mg of AFM was dissolved in 8M urea with 0.4 M ammonium bicarbonate, then reduced with DTT and carboxymethylated with iodoacetic acid. The sample was subsequently diluted with water to adjust the urea concentration to 2M, then digested with sequencing grade trypsin (Boehringer Mannheim) at 37°C for 18 h with an enzyme to substrate ratio of 1:50. The digested protein was injected on a Hewlett Packard 1090 HPLC equipped with a 4.6 x 250 mm C18 reverse phase column (Vydac). Elution was performed using a linear gradient of acetonitrile with 0.1 % trifluoroacetic acid at a flow rate of 0.75 ml/min. Elution was monitored at 214 nm and fractions were collected. Selected fractions were run on the Hewlett Packard G1004A protein sequencer as described above. The amino acid sequences obtained from select tryptic fragments are summarized in Table 1.

Table 1: Trypsin fragments

	FX18	YHYLIR	(SEQ ID NO:7)
5	FX20	FTFEYSR	(SEQ ID NO:8)
	FX27	FTDSENVQCERDADP	(SEQ ID NO:9)
10	FX29	IVQIYKDLLR	(SEQ ID NO:10)
	FX32	IAPQLSTEELVSLGE	(SEQ ID NO:11)
	FX37	RHPDLSIPELLR	(SEQ ID NO:12)
15	FX45	ESLLNHFLYEVAR	(SEQ ID NO:13)
	FX53	RNPVFVFAPTLLTVAVHFEEVAKSCC	(SEQ ID NO:14)

20 EXAMPLE 4: Isolation of AFM cDNA

The polymerase chain reaction (PCR) was used to amplify a portion of the AFM gene from which an exact probe could be derived. A PCR (PCR 1) was first performed with fully degenerate primers specifying the 25 sense strand for the N-terminal amino acid sequence QKFIEDN (SEQ ID NO: 15) [5' ACG CTG AAT TCG CCA (GA)AA (GA)TT (CT)AT (ATC)GA (GA)GA (CT)AA] (SEQ ID NO: 16) and the antisense strand for a portion of the FX 29 tryptic peptide sequence IVQIYKD (SEQ ID NO: 17) [5' ACG CTA 30 AGC TTG C(GA)T C(CT)T T(GA)T A(GAT)A T(CT)T G(AGCT)A C(GAT)A T] (SEQ ID NO: 18). One ng of Quick Clone Human liver cDNA (Clontech, cat. no. 7113-1) was used as the template in a 100 ml PCR performed in a standard buffer (Perkin-Elmer Cetus) with 1 mM of each degenerate 35 primer. The cycling parameters used in the PCR were as follows: 95°C , 8 min (1 cycle); 94°C, 1 min, 34°C, 10

min, 72°C, 2 min (3 cycles); 94°C, 1 min, 50°C, 1 min, 72°C, 2 min (45 cycles); 72°C, 5 min (1 cycle).

Agarose gel analysis of PCR 1 did not reveal the amplification of any specific products, thus we 5 utilized an aliquot of PCR 1 as a template for a second PCR (PCR 2) using a nested primer pair. For this experiment, we used fully degenerate primers specifying the sense strand for the N-terminal amino acid sequence DNIEYIT (SEQ ID NO: 19) [5' ACG CTG AAT TCG CGA (CT)AA 10 (CT)AT (ATC)GA (GA)TA (CT)AT (ATC)AC] (SEQ ID NO: 20) and the antisense strand for the FX 20 tryptic peptide sequence FTFEYS (SEQ ID NO: 21) [5' ACG CTA AGC TTG C(GATC)G A(GA)T A(CT)T C(GA)A A(ACGT)G T(GA)A A] (SEQ ID NO: 22). PCR 2 was then performed using the same 15 reaction mix and cycling parameters as PCR 1, except for the substitution of 1 ml of PCR 1 in place of the human liver cDNA. Analysis of PCR 2 by agarose gel electrophoresis revealed the amplification of a 1 kb product.

20 To prepare PCR 2 for DNA sequencing, the inventors utilized the *EcoR* I and *Hind* III sites that were incorporated into the degenerate primers. These sites were used to clone the fragment into mp19 (Boehringer Mannheim). The 1 kb PCR 2 product cloned in 25 mp19 (mp19 AFM) was then sequenced in its entirety from both strands. Nucleotide sequence analysis of the fragment confirmed that we had amplified a segment of AFM cDNA as the sequence was found to have an open reading frame which encoded 3 tryptic peptides that were 30 already sequenced (Table 1, Fxs 27, 45, 53). The nucleotide sequence of the 1 kb fragment was compared to all sequences in the Genbank database and found to be unique. We also observed that the partial AFM cDNA had significant homology with cDNAs reported for ALB family 35 proteins (ALB, AFP and VDB). Therefore, in order to isolate a full-length cDNA encoding AFM and to minimize

the probability of hybridizing other ALB family genes, we screened a human liver cDNA library with an exact 18mer oligonucleotide (5' TAT GTG CTA TGG AGG GGC) (SEQ ID NO: 23) derived from a segment of AFM sequence that 5 was not highly homologous to ALB, AFP and VDB. The oligonucleotide was end-labeled with $^{32}\text{P}_i$ and used to screen a human liver cDNA library (Clontech, cat. no. HL1115a). Positive clones were purified and rescreened with the same oligonucleotide probe. A single positive 10 clone (17AFM) with a 2.3 kb insert was chosen for further study and phage DNA was prepared. The 2.3 kb insert was then sequenced in its entirety from both strands and verified to encode AFM.

This approach enabled the inventors to isolate 15 a lambda phage (17AFM) containing full-length AFM cDNA.

EXAMPLE 5: Characterization of AFM

The insert in 17AFM is 2287 bp (FIG. 1) consisting of a 317 bp 5' untranslated region, a 1797 bp 20 sequence encoding a protein of 599 amino acids and a 173 bp 3' untranslated region. The predicted amino acid sequence of AFM was found to contain all the tryptic peptides (See Table 1) that had been previously sequenced from the purified protein. Translation of the 25 AFM cDNA sequence reveals that a 21 amino acid hydrophobic leader peptide precedes the experimentally determined N-terminus of mature AFM. The mature AFM is predicted to have 578 amino acids with a calculated Mr of 66576 and pI of 5.65. The difference between the 30 calculated Mr of AFM and its apparent molecular weight on SDS-PAGE is likely due to glycosylation. AFM has 4 potential sites for N-glycosylation (FIG. 1) and N-glycanase treatment reduced the apparent Mr of AFM to 65000 when analyzed by reducing SDS-PAGE (data not 35 shown).

A comparison of the deduced AFM amino acid sequence and other ALB family sequences is shown in FIG. 2. It is evident there is strong similarity between AFM and other ALB family members throughout the entire 5 protein. When conserved amino acids are taken into account, the of AFM to AFP, ALB and VDB is 60.4%, 54.8% and 41.2% respectively. The distribution of Cys residues is conserved among ALB family members. The positions of AFM Cys residues are clearly consistent with this 10 arrangement (FIG. 3). The Cys residues in ALB family proteins have also been proposed to form a pattern of disulfide bridges enabling these proteins to be depicted as a series of 9 double loops defining 3 structural 15 domains. FIG. 4 shows that the 34 Cys residues in AFM can be arranged into a pattern of 17 disulfide-linked pairs that parallels the domain organization observed in other ALB family proteins.

EXAMPLE 6: Chromosomal Mapping

20 PCR was performed on a panel of somatic cell hybrids (Bios Laboratories, cat. no. CP2-02) to identify the chromosomal location of the gene. For PCR, we utilized primers (5' CAA CCC TGC TGT GGA CCA C; 5' GCA CAT ATG TTT TAT CAG CTT T) (SEQ ID NO: 24 and SEQ ID NO: 25) that would be expected to amplify an 88 bp fragment between nt 1790 and 1878 in the AFM cDNA. Each PCR on 25 somatic cell hybrid DNA was performed in a standard 100 ml reaction mixture (Perkin-Elmer Cetus) containing 250 ng DNA and a final concentration of 0.1 mM of each 30 primer. The cycling parameters were as follows: 95°C, 5 min (1 cycle); 94°C, 1 min, 56°C, 1 min, 72°C, 1 min (25 cycles); 72°C, 5 min (1 cycle).

Utilizing PCR on a panel of commercially 35 available somatic cell hybrids (data not shown), we detected an amplified product in only 2 hybrids. Both of these hybrids had DNA in common from human chromosomes

4, 5 and 8. Since PCRs performed on hybrids containing DNA from chromosomes 5 and 8 did not yield amplified products, we conclude that AFM resides on human chromosome 4 along with other ALB family genes.

5

EXAMPLE 7: Stable Expression of rAFM

Two separate PCR's were performed on AFM cDNA to generate 2 overlapping fragments that span the entire AFM cDNA sequence. The oligonucleotide pair (5' TCA CCT 10 CTA GAC CAC CAT GAA ACT ACT AAA ACT TAC AG + 5' AAT TTC TCA GGA GAT CTT TGT ATA) (SEQ ID NO: 26 and SEQ ID NO: 27) used in the first PCR introduced an *Xba* I site and a perfect Kozak sequence preceeding the AFM initiator codon. The amplified product was subsequently cloned 15 into pGEMT (Promega) to create pDJ11. The oligonucleotide pair (5' AAA TAT ACA AAG ATC TCC TGA GAA + 5' TCC CGG TCG ACT CAG TTG CCA ATT TTT GGA C) (SEQ ID NO: 28 and SEQ ID NO: 29) used in the second PCR introduced a *Sal* I site following the natural stop codon 20 in AFM and the product was cloned into pGEMT to create pDJ13. The 3' end of AFM was then joined to the 5' end by ligating a *Bgl* II-*Sal* I fragment from pDJ13 into pDJ11 that had been digested with *Bgl* II and *Sal* I. The resultant plasmid (pDJ14) was then digested with *Xba* I 25 and *Sal* I and the entire AFM cDNA was cloned into the mammalian expression vector pDSRa (European Patent Application A20398753) that was modified to include unique *Xba* I and *Sal* I sites. The AFM expression vector was used to transfect a Chinese Hamster Ovary (CHO) cell 30 line deficient in dihydrofolate reductase (CHO D⁻). Transfectants were selected in medium lacking hypoxanthine and thymidine. An RNase protection assay was used to screen for transfectants that had AFM-specific mRNA. A single clone was grown without serum as 35 described (See, Bourdrel, L. et al., *Protein Expression*

Purif. 4: 130 (1993) to generate conditioned medium (CM) containing rAFM.

A single stable transfectant expressing high levels of AFM-specific mRNA was isolated. Immunoblots 5 performed with AM339 (see Example 12) demonstrates that this antibody recognizes both rAFM produced from the transfectant as well as natural AFM isolated from human plasma (FIG. 5). rAFM was purified from CM derived from this transfectant (FIG. 6) and SDS-PAGE demonstrated 10 rAFM to be greater than 95% pure with the same N- terminus as AFM purified from plasma (data not shown).

EXAMPLE 8: Protein Purification

Serum-free CM was concentrated 10-fold and 15 diafiltered against 25 mM sodium phosphate, pH 7.5 using a Filtron ultrafiltration apparatus loaded with 10 K molecular weight cut-off filters. The CM was then adjusted with 3.9 M ammonium sulfate to achieve a final ammonium sulfate concentration of 1.6 M. This mixture 20 was stirred for 0.5 hr; no precipitate was observed. The solution was then filtered in succession through 0.45mm and 0.22mm filters. The resultant filtrate was loaded onto a Phenyl Sepharose HP column previously 25 equilibrated with 50 mM sodium phosphate, 1.6 M ammonium sulfate; pH 7.5. Loading, washing and elution were done at a linear flow rate of 40 cm/hr. Following sample loading, the column was washed successively with 50 mM sodium phosphate, 1.6 M ammonium sulfate, pH 7.5 and 50 mM sodium phosphate, pH 7.5. rAFM was eluted with water 30 and detected by immunoblotting (see below).

The pH of the water eluate was adjusted to 8.0 with 0.1 M Tris-base and loaded at a flow rate of 100 cm/hr onto a Q Sepharose HP column previously equilibrated with 20 mM Tris-HCl, pH 8.0. Following 35 sample loading, the column was thoroughly washed with 20 mM Tris-HCl, pH 8.2, then with 0.1 M NaCl, 20 mM Tris-

HCl, pH 8.2. The column was then resolved with a 3 column volume (CV) linear gradient of NaCl ranging from 0 - 0.3 M, followed by a 1 CV linear gradient of NaCl ranging from 0.3 M to 1.0 M in 20 mM Tris-HCl, pH 8.0.

5 Fractions containing rAFM were pooled and loaded onto a Superdex 200 column previously equilibrated with phosphate-buffered saline pH 7.4. The column was loaded and resolved at a flow rate of 1 ml/min.

10 EXAMPLE 9: Delipidation of AFM

To one volume of protein solution was added 2.5 volumes of 1-butanol/diisopropyl ether (40:60). The mixture was shaken gently for 30 minutes at room temperature and then centrifuged for 5 minutes at 500 x g. The aqueous layer was recovered and to this was added a second 2.5 volumes of 1-butanol/diisopropyl ether (40:60). The sample was treated as before and the aqueous phase recovered. To the twice extracted aqueous phase was added one volume of diisopropyl ether, the mixture shaken and immediately centrifuged at 250 x g for 5 minutes. The aqueous layer was recovered and used as essentially delipidated.

EXAMPLE 10: Immunoblotting

25 All samples for immunoblotting were electrophoresed on 4-20% SDS polyacrylamide gels (Novex) and the proteins were then electrophoretically transferred to nitrocellulose membranes (Schleicher and Schuell, cat. no. BA83). The filters were first treated 30 with polyclonal antibody AM339 that was raised in rabbits against a synthetic peptide (DLSLREGKFTDSENV) (SEQ ID NO: 30) derived from amino acids 304 - 319 in AFM and then with donkey anti-rabbit Ig linked to horseradish peroxidase. Immune complexes were visualized 35 by enhanced chemiluminescence according to the manufacturer's (Amersham) specifications.

EXAMPLE 11: Nucleotide sequence comparisons

Nucleotide sequences were compared to the Genbank (Release 78.0) sequences using the method of Pearson and Lipman (Proc. Natl. Acad. Sci. USA 84, 2444-5 2448 (1988)) in the FASTA program of the Genetics Computer Group (GCG), Inc. (Madison, WI). Comparisons are shown in FIG. 2A.

10 EXAMPLE 12: Generation of peptide antibodies specific for AFM

Since AFM was similar to at least three other members of the albumin family, it was possible that antibodies generated against the entire protein would also cross react with other albumin family proteins. To 15 create AFM-specific antibodies, we first identified specific peptides of AFM which were dissimilar to peptide sequences found in the other albumin family proteins. These peptide sequences were then synthesized and the synthetic peptides were used to inject rabbits 20 to obtain polyclonal antisera against each peptide. The AFM peptides that were synthesized are as follows:

H1 EKLVKDMVEYKDRC (SEQ ID NO:31)

aa 43-56 in AFM

25 The corresponding antibody is referred to as AM384.

H2 CIINSNKDDRPKDLSLR (SEQ ID NO:32)

aa 292-308 in AFM

The corresponding antibody is referred to as AM 609.

30

H3 DLSLREGKFTDSENVNC (SEQ ID NO:30)

aa 304-319 in AFM

The corresponding antibody is referred to as AM 339.

H4 CQERDADPDTFFAKFT
aa 319-334 in AFM

(SEQ ID NO:33)

The corresponding antibody is referred to as AM 1104.

5 EXAMPLE 13: Generation of polyclonal antisera to AFM

Rabbit polyclonal antiserum was generated from purified AFM by methods that are standard in the art. The antiserum was found to bind specifically to AFM.

10 Discussion of Examples

ALB family proteins are comprised of three homologous folding domains and are predicted to have evolved from an ancestral gene that coded for an approximately 190 amino acid single domain protein
15 containing 3 double loops formed by 6 disulfide bridges. The genes in this family have all been mapped to the 4q11-q22 region of chromosome 4. AFM shares significant homology with ALB family proteins and has Cys residues consistent with a similar overall 3-domain organization.
20 In addition, AFM has been localized to chromosome 4. Thus, there is compelling evidence that AFM is the fourth member of the ALB family.

There are some noteworthy distinctions among ALB family members. Concentrations in adult serum are 50
25 ng/ml for AFP, 350 mg/ml for VDB, 40 mg/ml for ALB and 30 mg/ml for AFM (data obtained by immunoblot analysis). ALB is not glycosylated, AFP and VDB each have 1 potential N-glycosylation site while AFM has 4 potential sites. ALB expresses one free thiol group that has been
30 implicated in complex formation with Cys, glutathione, mercurial and gold compounds. In contrast, AFP and VDB have an even number of Cys residues and are thought not to have a free thiol. AFM has an even number of Cys residues, suggesting that it may not have a free thiol
35 and may not bind glutathione and mercurials as does ALB.

There also are differences in the intradomain disulfide bonding pattern among ALB family members. VDB is predicted to have a disulfide bridge in double loop 1A. This bridge is absent in ALB, AFP and AFM. A 5 disulfide bridge domain 2C is common to ALB, VDB and AFM but is not present in AFP. Thus, while the 4 ALB family proteins are evolutionarily related, there are clear differences in the molecular organization of these proteins.

10 Structural similarities between AFM and other ALB family members suggest that AFM could scavenge or transport a variety of ligands. We examined whether known ligand-binding sites in the sequence of ALB family proteins were also present in AFM. VDB has a binding 15 site for sterols between amino acids 35-49 and a binding site for actin between amino acids 373-403. Using the GCG GAP program and the alignment of FIG. 2A, AFM has 60% similarity and 40% identity between VDB amino acids 35-49 but only 32% similarity and 10% identity between 20 VDB amino acids 373-403. Thus, it is possible that AFM has sterol binding sites (e.g., the amino acids in AFM that correspond to amino acids 35 - 49 of VDB) but it is not likely to bind actin. The X-ray crystal structure 25 of ALB was used to show that ALB binds a variety of ligands (aspirin, warfarin, IS, DIS, TIB, bilirubin) between amino acids 186-260 in domain 2A and an array of ligands (aspirin, diazepam, digitoxin, clofibrate, ibuprofen, IS, DIS, TIB, long chain fatty acids) between amino acids 379-455 in domain 3A. A GCG GAP comparison 30 between analogous regions in AFM reveals that AFM has 54% similarity and 35% identity in domain 2A and 45% similarity and 25% identity in domain 3A. These moderate degrees of similarity make it possible but not conclusive as to whether AFM binds the same ligands as 35 ALB in domains 2A and 3A.

Abbreviations

The abbreviations used in the above Examples
5 are: AFM, afamin; AFP, a-fetoprotein; ALB, human serum
albumin; CHO, Chinese Hamster Ovary; CM, conditioned
medium; CV, column volume; DIS, 3, 5- diiodosalicyclic
acid; DTT, dithiothreitol; HPLC, high performance liquid
chromotography; IS, 5-iodosalicyclic acid; PAGE,
10 polyacrylamide gel electrophoresis; PBS, phosphate-
buffered saline; PCR, polymerase chain reaction; r,
recombinant; TIB, 2,3,5- triiodobenzoic acid; VDP,
vitamin D-binding protein.

15

* * *

The invention now being fully described, it
will be apparent to one of ordinary skill in the art
that many changes and modifications can be made thereto,
20 without departing from the spirit and scope of the
invention as set forth herein.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: AMGEN INC.

(ii) TITLE OF INVENTION: Afamin: A Human Serum Albumin-Like Protein

(iii) NUMBER OF SEQUENCES: 33

(iv) CORRESPONDENCE ADDRESS:

- (A) ADDRESSEE: Amgen Center, Patent Operations/RRC
- (B) STREET: 1840 DeHavilland Drive
- (C) CITY: Thousand Oaks
- (D) STATE: California
- (E) COUNTRY: U.S.
- (F) ZIP: 91320-1789

(v) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

- (A) APPLICATION NUMBER:
- (B) FILING DATE:
- (C) CLASSIFICATION:

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2287 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 318..2117

(ix) FEATURE:

- (A) NAME/KEY: mat_peptide
- (B) LOCATION: 381..2114

(ix) FEATURE:

- (A) NAME/KEY: sig_peptide
- (B) LOCATION: 318..380

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CCCCGAGTCT	CTGCGCCTTC	ACATAGTTGT	CACAGGACTA	AAGCAAATTG	ATCCAGGGGG	60
AAACACTGTA	GACCGTGTAT	ATAAAAACAC	TCTATAAACT	GCAATGCTCA	ATTCTTAGTA	120
TAACATATTGT	TGTTGTATTG	ATATTTATTA	GTATTGGTGC	TCACAAAAAG	AGTCTAAATT	180
CCATAAGTCT	TTATATTCAG	GCTACTCTT	ATTTTGAAA	ACTCATTTC	TATCACCTT	240
TTCTATTTA	CTCCATATTG	AGGCCTCATA	AATCCAATT	TTTATTTCTT	TCTTTGTAA	300
ATGTGGTTTC	TACAAAG	ATG AAA CTA CTA AAA CTT ACA GGT TTT ATT TTT				350
		Met Lys Leu Leu Lys Leu Thr Gly Phe Ile Phe				
	-21	-20		-15		
TTC TTG TTT TTT TTG ACT GAA TCC CTA ACC CTG CCC ACA CAA CCT CGG						398
Phe Leu Phe Phe Leu Thr Glu Ser Leu Thr Leu Pro Thr Gln Pro Arg						
-10	-5		1		5	
GAT ATA GAG AAC TTC AAT AGT ACT CAA AAA TTT ATA GAA GAT AAT ATT						446
Asp Ile Glu Asn Phe Asn Ser Thr Gln Lys Phe Ile Glu Asp Asn Ile						
10	15		20			
GAA TAC ATC ACC ATC ATT GCA TTT GCT CAG TAT GTT CAG GAA GCA ACC						494
Glu Tyr Ile Thr Ile Ile Ala Phe Ala Gln Tyr Val Gln Glu Ala Thr						
25	30		35			
TTT GAA GAA ATG GAA AAG CTG GTG AAA GAC ATG GTA GAA TAC AAA GAC						542
Phe Glu Glu Met Glu Lys Leu Val Lys Asp Met Val Glu Tyr Lys Asp						
40	45		50			
AGA TGT ATG GCT GAC AAG ACG CTC CCA GAG TGT TCA AAA TTA CCT AAT						590
Arg Cys Met Ala Asp Lys Thr Leu Pro Glu Cys Ser Lys Leu Pro Asn						
55	60		65		70	
AAT GTT TTA CAG GAA AAA ATA TGT GCT ATG GAG GGG CTG CCA CAA AAG						638
Asn Val Leu Gln Glu Lys Ile Cys Ala Met Glu Gly Leu Pro Gln Lys						
75	80		85			
CAT AAT TTC TCA CAC TGC TGC AGT AAG GTT GAT GCT CAA AGA AGA CTC						686
His Asn Phe Ser His Cys Cys Ser Lys Val Asp Ala Gln Arg Arg Leu						
90	95		100			
TGT TTC TTC TAT AAC AAG AAA TCT GAT GTG GGA TTT CTG CCT CCT TTC						734
Cys Phe Phe Tyr Asn Lys Lys Ser Asp Val Gly Phe Leu Pro Pro Phe						
105	110		115			
CCT ACC CTG GAT CCC GAA GAG AAA TGC CAG GCT TAT GAA AGT AAC AGA						782
Pro Thr Leu Asp Pro Glu Glu Lys Cys Gln Ala Tyr Glu Ser Asn Arg						
120	125		130			
GAA TCC CTT TTA AAT CAC TTT TTA TAT GAA GTT GCC AGA AGG AAC CCA						830
Glu Ser Leu Leu Asn His Phe Leu Tyr Glu Val Ala Arg Arg Asn Pro						
135	140		145		150	
TTT GTC TTC GCC CCT ACA CTT CTA ACT GTT GCT GTT CAT TTT GAG GAG						878

Phe Val Phe Ala Pro Thr Leu Leu Thr Val Ala Val His Phe Glu Glu
 155 160 165
 GTG GCC AAA TCA TGT TGT GAA GAA CAA AAC AAA GTC AAC TGC CTT CAA 926
 Val Ala Lys Ser Cys Cys Glu Glu Gln Asn Lys Val Asn Cys Leu Gln
 170 175 180
 ACA AGG GCA ATA CCT GTC ACA CAA TAT TTA AAA GCA TTT TCT TCT TAT 974
 Thr Arg Ala Ile Pro Val Thr Gln Tyr Leu Lys Ala Phe Ser Ser Tyr
 185 190 195
 CAA AAA CAT GTC TGT GGG GCA CTT TTG AAA TTT GGA ACC AAA GTT GTA 1022
 Gln Lys His Val Cys Gly Ala Leu Leu Lys Phe Gly Thr Lys Val Val
 200 205 210
 CAC TTT ATA TAT ATT GCG ATA CTC AGT CAA AAA TTC CCC AAG ATT GAA 1070
 His Phe Ile Tyr Ile Ala Ile Leu Ser Gln Lys Phe Pro Lys Ile Glu
 215 220 225 230
 TTT AAG GAG CTT ATT TCT CTT GTA GAA GAT GTT TCT TCC AAC TAT GAT 1118
 Phe Lys Glu Leu Ile Ser Leu Val Glu Asp Val Ser Ser Asn Tyr Asp
 235 240 245
 GGA TGC TGT GAA GGG GAT GTT GTG CAG TGC ATC CGT GAC ACG AGC AAG 1166
 Gly Cys Cys Glu Gly Asp Val Val Gln Cys Ile Arg Asp Thr Ser Lys
 250 255 260
 GTT ATG AAC CAT ATT TGT TCA AAA CAA GAT TCT ATC TCC AGC AAA ATC 1214
 Val Met Asn His Ile Cys Ser Lys Gln Asp Ser Ile Ser Ser Lys Ile
 265 270 275
 AAA GAG TGC TGT GAA AAG AAA ATA CCA GAG CGC GGC CAG TGC ATA ATT 1262
 Lys Glu Cys Cys Glu Lys Ile Pro Glu Arg Gly Gln Cys Ile Ile
 280 285 290
 AAC TCA AAC AAA GAT GAT AGA CCA AAG GAT TTA TCT CTA AGA GAA GGA 1310
 Asn Ser Asn Lys Asp Asp Arg Pro Lys Asp Leu Ser Leu Arg Glu Gly
 295 300 305 310
 AAA TTT ACT GAC AGT GAA AAT GTG TGT CAA GAA CGA GAT GCT GAC CCA 1358
 Lys Phe Thr Asp Ser Glu Asn Val Cys Gln Glu Arg Asp Ala Asp Pro
 315 320 325
 GAC ACC TTC TTT GCG AAG TTT ACT TTT GAA TAC TCA AGG AGA CAT CCA 1406
 Asp Thr Phe Ala Lys Phe Thr Phe Glu Tyr Ser Arg Arg His Pro
 330 335 340
 GAC CTG TCT ATA CCA GAG CTT TTA AGA ATT GTT CAA ATA TAC AAA GAT 1454
 Asp Leu Ser Ile Pro Glu Leu Leu Arg Ile Val Gln Ile Tyr Lys Asp
 345 350 355
 CTC CTG AGA AAT TGC TGC AAC ACA GAA AAC CCT CCA GGT TGT TAC CGT 1502
 Leu Leu Arg Asn Cys Cys Asn Thr Glu Asn Pro Pro Gly Cys Tyr Arg
 360 365 370
 TAC GCG GAA GAC AAA TTC AAT GAG ACA ACT GAG AAA AGC CTC AAG ATG 1550
 Tyr Ala Glu Asp Lys Phe Asn Glu Thr Thr Glu Lys Ser Leu Lys Met
 375 380 385 390

GTA CAA CAA GAA TGT AAA CAT TTC CAG AAT TTG GGG AAG GAT GGT TTG Val Gln Gln Glu Cys Lys His Phe Gln Asn Leu Gly Lys Asp Gly Leu 395 400 405	1598
AAA TAC CAT TAC CTC ATC AGG CTC ACG AAG ATA GCT CCC CAA CTC TCC Lys Tyr His Tyr Leu Ile Arg Leu Thr Lys Ile Ala Pro Gln Leu Ser 410 415 420	1646
ACT GAA GAA CTG GTG TCT CTT GGC GAG AAA ATG GTG ACA GCT TTC ACT Thr Glu Glu Leu Val Ser Leu Gly Glu Lys Met Val Thr Ala Phe Thr 425 430 435	1694
ACT TGC TGT ACG CTA AGT GAA GAG TTT GCC TGT GTT GAT AAT TTG GCA Thr Cys Cys Thr Leu Ser Glu Glu Phe Ala Cys Val Asp Asn Leu Ala 440 445 450	1742
GAT TTA GTT TTT GGA GAG TTA TGT GGA GTA AAT GAA AAT CGA ACT ATC Asp Leu Val Phe Gly Glu Leu Cys Gly Val Asn Glu Asn Arg Thr Ile 455 460 465 470	1790
AAC CCT GCT GTG GAC CAC TGC TGT AAA ACA AAC TTT GCC TTC AGA AGG Asn Pro Ala Val Asp His Cys Cys Lys Thr Asn Phe Ala Phe Arg Arg 475 480 485	1838
CCC TGC TTT GAG AGT TTG AAA GCT GAT AAA ACA TAT GTG CCT CCA CCT Pro Cys Phe Glu Ser Leu Lys Ala Asp Lys Thr Tyr Val Pro Pro Pro 490 495 500	1886
TTC TCT CAA GAT TTA TTT ACC TTT CAC GCA GAC ATG TGT CAA TCT CAG Phe Ser Gln Asp Leu Phe Thr Phe His Ala Asp Met Cys Gln Ser Gln 505 510 515	1934
AAT GAG GAG CTT CAG AGG AAG ACA GAC AGG TTT CTT GTC AAC TTA GTG Asn Glu Glu Leu Gln Arg Lys Thr Asp Arg Phe Leu Val Asn Leu Val 520 525 530	1982
AAG CTG AAG CAT GAA CTC ACA GAT GAA GAG CTG CAG TCT TTG TTT ACA Lys Leu Lys His Glu Leu Thr Asp Glu Glu Leu Gln Ser Leu Phe Thr 535 540 545 550	2030
AAT TTC GCA AAT GTA GTG GAT AAG TGC TGC AAA GCA GAG AGT CCT GAA Asn Phe Ala Asn Val Val Asp Lys Cys Cys Lys Ala Glu Ser Pro Glu 555 560 565	2078
GTC TGC TTT AAT GAA GAG AGT CCA AAA ATT GGC AAC TGAAGCCAGC Val Cys Phe Asn Glu Glu Ser Pro Lys Ile Gly Asn 570 575	2124
TGCTGGAGAT ATGTAAAGAA AAAAGCACCA AAGGGAGGC TTCCTATCTG TGTGGTGATG	2184
AATCGCATT CCTGAGAACAA AAATAAAAGG ATTTTCTGT AACTGTCACC TGAAATAATA	2244
CATTGCAGCA AGCAATAAAC ACAACATTGTAA AAA	2287

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 599 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Lys Leu Leu Lys Leu Thr Gly Phe Ile Phe Phe Leu Phe Phe Leu
-21 -20 -15 -10

Thr Glu Ser Leu Thr Leu Pro Thr Gln Pro Arg Asp Ile Glu Asn Phe
-5 1 5 10

Asn Ser Thr Gln Lys Phe Ile Glu Asp Asn Ile Glu Tyr Ile Thr Ile
15 20 25

Ile Ala Phe Ala Gln Tyr Val Gln Glu Ala Thr Phe Glu Glu Met Glu
30 35 40

Lys Leu Val Lys Asp Met Val Glu Tyr Lys Asp Arg Cys Met Ala Asp
45 50 55

Lys Thr Leu Pro Glu Cys Ser Lys Leu Pro Asn Asn Val Leu Gln Glu
60 65 70 75

Lys Ile Cys Ala Met Glu Gly Leu Pro Gln Lys His Asn Phe Ser His
80 85 90

Cys Cys Ser Lys Val Asp Ala Gln Arg Arg Leu Cys Phe Phe Tyr Asn
95 100 105

Lys Lys Ser Asp Val Gly Phe Leu Pro Pro Phe Pro Thr Leu Asp Pro
110 115 120

Glu Glu Lys Cys Gln Ala Tyr Glu Ser Asn Arg Glu Ser Leu Leu Asn
125 130 135

His Phe Leu Tyr Glu Val Ala Arg Arg Asn Pro Phe Val Phe Ala Pro
140 145 150 155

Thr Leu Leu Thr Val Ala Val His Phe Glu Glu Val Ala Lys Ser Cys
160 165 170

Cys Glu Glu Gln Asn Lys Val Asn Cys Leu Gln Thr Arg Ala Ile Pro
175 180 185

Val Thr Gln Tyr Leu Lys Ala Phe Ser Ser Tyr Gln Lys His Val Cys
190 195 200

Gly Ala Leu Leu Lys Phe Gly Thr Lys Val Val His Phe Ile Tyr Ile
205 210 215

Ala Ile Leu Ser Gln Lys Phe Pro Lys Ile Glu Phe Lys Glu Leu Ile
220 225 230 235

Ser Leu Val Glu Asp Val Ser Ser Asn Tyr Asp Gly Cys Cys Glu Gly

240 245 250

Asp Val Val Gln Cys Ile Arg Asp Thr Ser Lys Val Met Asn His Ile
255 260 265

Cys Ser Lys Gln Asp Ser Ile Ser Ser Lys Ile Lys Glu Cys Cys Glu
270 275 280

Lys Lys Ile Pro Glu Arg Gly Gln Cys Ile Ile Asn Ser Asn Lys Asp
285 290 295

Asp Arg Pro Lys Asp Leu Ser Leu Arg Glu Gly Lys Phe Thr Asp Ser
300 305 310 315

Glu Asn Val Cys Gln Glu Arg Asp Ala Asp Pro Asp Thr Phe Phe Ala
320 325 330

Lys Phe Thr Phe Glu Tyr Ser Arg Arg His Pro Asp Leu Ser Ile Pro
335 340 345

Glu Leu Leu Arg Ile Val Gln Ile Tyr Lys Asp Leu Leu Arg Asn Cys
350 355 360

Cys Asn Thr Glu Asn Pro Pro Gly Cys Tyr Arg Tyr Ala Glu Asp Lys
365 370 375

Phe Asn Glu Thr Thr Glu Lys Ser Leu Lys Met Val Gln Gln Glu Cys
380 385 390 395

Lys His Phe Gln Asn Leu Gly Lys Asp Gly Leu Lys Tyr His Tyr Leu
400 405 410

Ile Arg Leu Thr Lys Ile Ala Pro Gln Leu Ser Thr Glu Glu Leu Val
415 420 425

Ser Leu Gly Glu Lys Met Val Thr Ala Phe Thr Thr Cys Cys Thr Leu
430 435 440

Ser Glu Glu Phe Ala Cys Val Asp Asn Leu Ala Asp Leu Val Phe Gly
445 450 455

Glu Leu Cys Gly Val Asn Glu Asn Arg Thr Ile Asn Pro Ala Val Asp
460 465 470 475

His Cys Cys Lys Thr Asn Phe Ala Phe Arg Arg Pro Cys Phe Glu Ser
480 485 490

Leu Lys Ala Asp Lys Thr Tyr Val Pro Pro Pro Phe Ser Gln Asp Leu
495 500 505

Phe Thr Phe His Ala Asp Met Cys Gln Ser Gln Asn Glu Glu Leu Gln
510 515 520

Arg Lys Thr Asp Arg Phe Leu Val Asn Leu Val Lys Leu Lys His Glu
525 530 535

Leu Thr Asp Glu Glu Leu Gln Ser Leu Phe Thr Asn Phe Ala Asn Val
540 545 550 555

Val Asp Lys Cys Cys Lys Ala Glu Ser Pro Glu Val Cys Phe Asn Glu
560 565 570

Glu Ser Pro Lys Ile Gly Asn
575

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 609 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Lys Trp Val Thr Phe Ile Ser Leu Leu Phe Leu Phe Ser Ser Ala
1 5 10 15

Tyr Ser Arg Gly Val Phe Arg Arg Asp Ala His Lys Ser Glu Val Ala
20 25 30

His Arg Phe Lys Asp Leu Gly Glu Glu Asn Phe Lys Ala Leu Val Leu
35 40 45

Ile Ala Phe Ala Gln Tyr Leu Gln Gln Cys Pro Phe Glu Asp His Val
50 55 60

Lys Leu Val Asn Glu Val Thr Glu Phe Ala Lys Thr Cys Val Ala Asp
65 70 75 80

Glu Ser Ala Glu Asn Cys Asp Lys Ser Leu His Thr Leu Phe Gly Asp
85 90 95

Lys Leu Cys Thr Val Ala Thr Leu Arg Glu Thr Tyr Gly Glu Met Ala
100 105 110

Asp Cys Cys Ala Lys Gln Glu Pro Glu Arg Asn Glu Cys Phe Leu Gln
115 120 125

His Lys Asp Asp Asn Pro Asn Leu Pro Arg Leu Val Arg Pro Glu Val
130 135 140

Asp Val Met Cys Thr Ala Phe His Asp Asn Glu Glu Thr Phe Leu Lys
145 150 155 160

Lys Tyr Leu Tyr Glu Ile Ala Arg Arg His Pro Tyr Phe Tyr Ala Pro
165 170 175

Glu Leu Leu Phe Phe Ala Lys Arg Tyr Lys Ala Ala Phe Thr Glu Cys
180 185 190

Cys Gln Ala Ala Asp Lys Ala Ala Cys Leu Leu Pro Lys Leu Asp Glu
195 200 205

Leu Arg Asp Glu Gly Lys Ala Ser Ser Ala Lys Gln Arg Leu Lys Cys
210 215 220

Ala Ser Leu Gln Lys Phe Gly Glu Arg Ala Phe Lys Ala Trp Ala Val
225 230 235 240

Ala Arg Leu Ser Gln Arg Phe Pro Lys Ala Glu Phe Ala Glu Val Ser
245 250 255

Lys Leu Val Thr Asp Leu Thr Lys Val His Thr Glu Cys Cys His Gly
260 265 270

Asp Leu Leu Glu Cys Ala Asp Asp Arg Ala Asp Leu Ala Lys Tyr Ile
275 280 285

Cys Glu Asn Gln Asp Ser Ile Ser Ser Lys Leu Lys Glu Cys Cys Glu
290 295 300

Lys Pro Leu Leu Glu Lys Ser His Cys Ile Ala Glu Val Glu Asn Asp
305 310 315 320

Glu Met Pro Ala Asp Leu Pro Ser Leu Ala Ala Asp Phe Val Glu Ser
325 330 335

Lys Asp Val Cys Lys Asn Tyr Ala Glu Ala Lys Asp Val Phe Leu Gly
340 345 350

Met Phe Leu Tyr Glu Tyr Ala Arg Arg His Pro Asp Tyr Ser Val Val
355 360 365

Leu Leu Leu Arg Leu Ala Lys Thr Tyr Glu Thr Thr Leu Glu Lys Cys
370 375 380

Cys Ala Ala Ala Asp Pro His Glu Cys Tyr Ala Lys Val Phe Asp Glu
385 390 395 400

Phe Lys Pro Leu Val Glu Glu Pro Gln Asn Leu Ile Lys Gln Asn Cys
405 410 415

Glu Leu Phe Lys Gln Leu Gly Glu Tyr Lys Phe Gln Asn Ala Leu Leu
420 425 430

Val Arg Tyr Thr Lys Lys Val Pro Gln Val Ser Thr Pro Thr Leu Val
435 440 445

Glu Val Ser Arg Asn Leu Gly Lys Val Gly Ser Lys Cys Cys Lys His
450 455 460

Pro Glu Ala Lys Arg Met Pro Cys Ala Glu Asp Tyr Leu Ser Val Val
465 470 475 480

Leu Asn Gln Leu Cys Val Leu His Glu Lys Thr Pro Val Ser Asp Arg
485 490 495

Val Thr Lys Cys Cys Thr Glu Ser Leu Val Asn Arg Arg Pro Cys Phe

	500	505	510
Ser Ala Leu Glu Val Asp Glu Thr Tyr Val Pro Lys Glu Phe Asn Ala			
515	520	525	
Glu Thr Phe Thr Phe His Ala Asp Ile Cys Thr Leu Ser Glu Lys Glu			
530	535	540	
Arg Gln Ile Lys Lys Gln Thr Ala Leu Val Glu Leu Val Lys His Lys			
545	550	555	560
Pro Lys Ala Thr Lys Glu Gln Leu Lys Ala Val Met Asp Asp Phe Ala			
565	570	575	
Ala Phe Val Glu Lys Cys Cys Lys Ala Asp Asp Lys Glu Thr Cys Phe			
580	585	590	
Ala Glu Glu Gly Lys Lys Leu Val Ala Ala Ser Gln Ala Ala Leu Gly			
595	600	605	

Leu

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 609 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Lys Trp Val Glu Ser Ile Phe Leu Ile Phe Leu Leu Asn Phe Thr			
1	5	10	15
Glu Ser Arg Thr Leu His Arg Asn Glu Tyr Gly Ile Ala Ser Ile Leu			
20	25	30	
Asp Ser Tyr Gln Cys Thr Ala Glu Ile Ser Leu Ala Asp Leu Ala Thr			
35	40	45	
Ile Phe Phe Ala Gln Phe Val Gln Glu Ala Thr Tyr Lys Glu Val Ser			
50	55	60	
Lys Met Val Lys Asp Ala Leu Thr Ala Ile Glu Lys Pro Thr Gly Asp			
65	70	75	80
Glu Gln Ser Ser Gly Cys Leu Glu Asn Gln Leu Pro Ala Phe Leu Glu			
85	90	95	
Glu Leu Cys His Glu Lys Glu Ile Leu Glu Lys Tyr Gly His Ser Asp			
100	105	110	

Cys Cys Ser Gln Ser Glu Glu Gly Arg His Asn Cys Phe Leu Ala His
115 120 125

Lys Lys Pro Thr Pro Ala Ser Ile Pro Leu Phe Gln Val Pro Glu Pro
130 135 140

Val Thr Ser Cys Glu Ala Tyr Glu Glu Asp Arg Glu Thr Phe Met Asn
145 150 155 160

Lys Phe Ile Tyr Glu Ile Ala Arg Arg His Pro Phe Leu Tyr Ala Pro
165 170 175

Thr Ile Leu Leu Trp Ala Ala Arg Tyr Asp Lys Ile Ile Pro Ser Cys
180 185 190

Cys Lys Ala Glu Asn Ala Val Glu Cys Phe Gln Thr Lys Ala Ala Thr
195 200 205

Val Thr Lys Glu Leu Arg Glu Ser Ser Leu Leu Asn Gln His Ala Cys
210 215 220

Ala Val Met Lys Asn Phe Gly Thr Arg Thr Phe Gln Ala Ile Thr Val
225 230 235 240

Thr Lys Leu Ser Gln Lys Phe Thr Lys Val Asn Phe Thr Glu Ile Gln
245 250 255

Lys Leu Val Leu Asp Val Ala His Val His Glu His Cys Cys Arg Gly
260 265 270

Asp Val Leu Asp Cys Leu Gln Asp Gly Glu Lys Ile Met Ser Tyr Ile
275 280 285

Cys Ser Gln Gln Asp Thr Leu Ser Asn Lys Ile Thr Glu Cys Cys Lys
290 295 300

Leu Thr Thr Leu Glu Arg Gly Gln Cys Ile Ile His Ala Glu Asn Asp
305 310 315 320

Glu Lys Pro Glu Gly Leu Ser Pro Asn Leu Asn Arg Phe Leu Gly Asp
325 330 335

Arg Asp Phe Asn Gln Phe Ser Ser Gly Glu Lys Asn Ile Phe Leu Ala
340 345 350

Ser Phe Val His Glu Tyr Ser Arg Arg His Pro Gln Leu Ala Val Ser
355 360 365

Val Ile Leu Arg Val Ala Lys Gly Tyr Gln Glu Leu Leu Glu Lys Cys
370 375 380

Phe Gln Thr Glu Asn Pro Leu Glu Cys Gln Asp Lys Gly Glu Glu Glu
385 390 395 400

Leu Gln Lys Tyr Ile Gln Glu Ser Gln Ala Leu Ala Lys Arg Ser Cys
405 410 415

Gly Leu Phe Gln Lys Leu Gly Glu Tyr Tyr Leu Gln Asn Ala Phe Leu

420 425 430

Val Ala Tyr Thr Lys Lys Ala Pro Gln Leu Thr Ser Ser Glu Leu Met
435 440 445

Ala Ile Thr Arg Lys Met Ala Ala Thr Ala Ala Thr Cys Cys Gln Leu
450 455 460

Ser Glu Asp Lys Leu Leu Ala Cys Gly Glu Gly Ala Ala Asp Ile Ile
465 470 475 480

Ile Gly His Leu Cys Ile Arg His Glu Met Thr Pro Val Asn Pro Gly
485 490 495

Val Gly Gln Cys Cys Thr Ser Ser Tyr Ala Asn Arg Arg Pro Cys Phe
500 505 510

Ser Ser Leu Val Val Asp Glu Thr Tyr Val Pro Pro Ala Phe Ser Asp
515 520 525

Asp Lys Phe Ile Phe His Lys Asp Leu Cys Gln Ala Gln Gly Val Ala
530 535 540

Leu Gln Thr Met Lys Gln Glu Phe Leu Ile Asn Leu Val Lys Gln Lys
545 550 555 560

Pro Gln Ile Thr Glu Glu Gln Leu Glu Ala Val Ile Ala Asp Phe Ser
565 570 575

Gly Leu Leu Glu Lys Cys Cys Gln Gly Gln Glu Gln Glu Val Cys Phe
580 585 590

Ala Glu Glu Gly Gln Lys Leu Ile Ser Lys Thr Arg Ala Ala Leu Gly
595 600 605

Val

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 474 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Lys Arg Val Leu Val Leu Leu Ala Val Ala Phe Gly His Ala
1 5 10 15

Leu Glu Arg Gly Arg Asp Tyr Glu Lys Asn Lys Val Cys Lys Glu Phe
20 25 30

Ser His Leu Gly Lys Glu Asp Phe Thr Ser Leu Ser Leu Val Leu Tyr
35 40 45

Ser Arg Lys Phe Pro Ser Gly Thr Phe Glu Gln Val Ser Gln Leu Val
50 55 60

Lys Glu Val Val Ser Leu Thr Glu Ala Cys Cys Ala Glu Gly Ala Asp
65 70 75 80

Pro Asp Cys Tyr Asp Thr Arg Thr Ser Ala Leu Ser Ala Lys Ser Cys
85 90 95

Glu Ser Asn Ser Pro Phe Pro Val His Pro Gly Thr Ala Glu Cys Cys
100 105 110

Thr Lys Glu Gly Leu Glu Arg Lys Leu Cys Met Ala Ala Leu Lys His
115 120 125

Gln Pro Gln Glu Phe Pro Thr Tyr Val Glu Pro Thr Asn Asp Glu Ile
130 135 140

Cys Glu Ala Phe Arg Lys Asp Pro Lys Glu Tyr Ala Asn Gln Phe Met
145 150 155 160

Trp Glu Tyr Ser Thr Asn Tyr Gly Gln Ala Pro Leu Ser Leu Leu Val
165 170 175

Ser Tyr Thr Lys Ser Tyr Leu Ser Met Val Gly Ser Cys Cys Thr Ser
180 185 190

Ala Ser Pro Thr Val Cys Phe Leu Lys Glu Arg Leu Gln Leu Lys His
195 200 205

Leu Ser Leu Leu Thr Thr Leu Ser Asn Arg Val Cys Ser Gln Tyr Ala
210 215 220

Ala Tyr Gly Glu Lys Lys Ser Arg Leu Ser Asn Leu Ile Lys Leu Ala
225 230 235 240

Gln Lys Val Pro Thr Ala Asp Leu Glu Asp Val Leu Pro Leu Ala Glu
245 250 255

Asp Ile Thr Asn Ile Leu Ser Lys Cys Cys Glu Ser Ala Ser Glu Asp
260 265 270

Cys Met Ala Lys Glu Leu Pro Glu His Thr Val Lys Leu Cys Asp Asn
275 280 285

Leu Ser Thr Lys Asn Ser Lys Phe Glu Asp Cys Cys Gln Glu Lys Thr
290 295 300

Ala Met Asp Val Phe Val Cys Thr Tyr Phe Met Pro Ala Ala Gln Leu
305 310 315 320

Pro Glu Leu Pro Asp Val Glu Leu Pro Thr Asn Lys Asp Val Cys Asp
325 330 335

Pro Gly Asn Thr Lys Val Met Asp Lys Tyr Thr Phe Glu Leu Ser Arg

340 345 350

Arg Thr His Leu Pro Glu Val Phe Leu Ser Lys Val Leu Glu Pro Thr
355 360 365

Leu Lys Ser Leu Gly Glu Cys Cys Asp Val Glu Asp Ser Thr Thr Cys
370 375 380

Phe Asn Ala Lys Gly Pro Leu Leu Lys Lys Glu Leu Ser Ser Phe Ile
385 390 395 400

Asp Lys Gly Gln Glu Leu Cys Ala Asp Tyr Ser Glu Asn Thr Phe Thr
405 410 415

Glu Tyr Lys Lys Lys Leu Ala Glu Arg Leu Lys Ala Lys Leu Pro Asp
420 425 430

Ala Thr Pro Lys Glu Leu Ala Lys Leu Val Asn Lys Arg Ser Asp Phe
435 440 445

Ala Ser Asn Cys Cys Ser Ile Asn Ser Pro Pro Leu Tyr Cys Asp Ser
450 455 460

Glu Ile Asp Ala Glu Leu Lys Asn Ile Leu
465 470

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 35 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Leu Pro Thr Gln Pro Arg Asp Ile Glu Asn Phe Xaa Ser Thr Gln Lys
1 5 10 15

Phe Ile Glu Asp Asn Ile Glu Tyr Ile Thr Ile Ile Ala Phe Ala Gln
20 25 30

Tyr Val Gln
35

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 6 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Tyr His Tyr Leu Ile Arg
1 5

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Phe Thr Phe Glu Tyr Ser Arg
1 5

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Phe Thr Asp Ser Glu Asn Val Cys Gln Glu Arg Asp Ala Asp Pro
1 5 10 15

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 10 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Ile Val Gln Ile Tyr Lys Asp Leu Leu Arg
1 5 10

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Ile Ala Pro Gln Leu Ser Thr Glu Glu Leu Val Ser Leu Gly Glu
1 5 10 15

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 12 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Arg His Pro Asp Leu Ser Ile Pro Glu Leu Leu Arg
1 5 10

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 13 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Glu Ser Leu Leu Asn His Phe Leu Tyr Glu Val Ala Arg
1 5 10

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown

(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Arg Asn Pro Phe Val Phe Ala Pro Thr Leu Leu Thr Val Ala Val His
1 5 10 15

Phe Glu Glu Val Ala Lys Ser Cys Cys
20 25

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Gln Lys Phe Ile Glu Asp Asn
1 5

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

ACGCTGAATT CGCCARAART TYATHGARGA YAA

33

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Ile Val Gln Ile Tyr Lys Asp
1 5

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 34 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

ACGCTAAGCT TGCRTCYTTR TADATYTGNA CDAT

34

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Asp Asn Ile Glu Tyr Ile Thr
1 5

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

ACGCTGAATT CGCGAYAAYA THGARTAYAT HAC

33

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 6 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Phe Thr Phe Glu Tyr Ser
1 5

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 31 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

ACGCTAAGCT TGCNGARTAY TCRAANGTRA A

31

(2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

TATGTGCTAT GGAGGGGC

18

(2) INFORMATION FOR SEQ ID NO:24:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

CAACCCTGCT GTGGACCA

18

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 22 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

GCACATATGT TTTATCAGCT TT

22

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 38 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

TCACCTCTAG ACCACCATGA AACTACTAAA ACTTACAG

38

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

AATTTCTCAG GAGATCTTG TATA

24

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown

(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

AAATATACAA AGATCTCCTG AGAA

24

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 31 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

TCCCGGTCGA CTCAGTTGCC AATTTTGGA C

31

(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Asp Leu Ser Leu Arg Glu Gly Lys Phe Thr Asp Ser Glu Asn Val Cys
1 5 10 15

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 14 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

Glu Lys Leu Val Lys Asp Met Val Glu Tyr Lys Asp Arg Cys
1 5 10

(2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 17 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Cys Ile Ile Asn Ser Asn Lys Asp Asp Arg Pro Lys Asp Leu Ser Leu
1 5 10 15
Arg

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Cys Gln Glu Arg Asp Ala Asp Pro Asp Thr Phe Phe Ala Lys Phe Thr
1 5 10 15

The embodiments of the invention in which an exclusive property or privilege is claimed are defined as follows:

5 1. A purified and isolated polynucleotide encoding AFM polypeptide or a variant thereof possessing a biological activity specific to AFM.

10 2. The polynucleotide of claim 1 which is a DNA sequence.

 3. The DNA sequence according to claim 2 which is a cDNA sequence or a biological replica thereof.

15 4. The cDNA sequence of claim 3, which encodes AFM.

 5. The DNA sequence according to claim 3 which is a genomic DNA sequence or a biological replica thereof.

20 6. The DNA sequence of claim 3 further including an endogenous expression control DNA sequence.

25 7. The DNA sequence according to claim 3 which is a wholly or partially chemically synthesized DNA sequence or a biological replica thereof.

 8. A DNA vector comprising a DNA sequence according to claim 3.

30 9. The vector of claim 8 wherein said DNA sequence is operatively linked to an expression control DNA sequence.

10. A host cell stably transformed or transfected with a DNA sequence according to claim 3 in a manner allowing the expression in said host cell of AFM 5 polypeptide or a variant thereof possessing a biological activity specific to AFM.

11. A method for producing AFM polypeptide or a variant thereof possessing a biological activity specific 10 to AFM, said method comprising growing a host cell according to claim 8 in a suitable nutrient medium and isolating AFM polypeptide or variant thereof from said cell or the growth medium.

15 12. Purified and isolated AFM polypeptide or a variant thereof possessing a biological activity specific to AFM.

20 13. The polypeptide of claim 12 comprising amino acid residues 1 through 578 of FIG 2.

14. The polypeptide of claim 12 comprising residues -23 through 578 of FIG 2.

25 15. Purified and isolated AFM polypeptide complexed with Apolipoprotein A1.

16. An antibody specific for AFM or a variant thereof.

30 17. A monoclonal antibody according to claim 16.

18. A humanized antibody according to claims 16 or 17.

19. A method for modulating a biological 5 activity of AFM comprising contacting AFM with an antibody according to claim 16 or 17.

20. A method for detecting the capacity of a cell to synthesize AFM comprising hybridizing a detectable 10 polynucleotide encoding AFM or a fragment thereof with RNA of said cell.

21. A method for detecting the capacity of a cell to synthesize AFM comprising reacting an antibody 15 according to claim 16 with polypeptides produced by said cell.

22. A method for determining the presence of AFM in a sample body fluid, comprising contacting the body 20 fluid with an antibody according to Claim 16.

23. An antisense polynucleotide for a polynucleotide encoding AFM.

25 24. An antisense polynucleotide for a DNA specifying an endogenous expression control DNA sequence of AFM.

30 25. A hybrid fusion polypeptide comprising, at its amino terminal, an AFM polypeptide or a variant thereof possessing a biological activity specific to AFM and, at its carboxy terminal, at least one constant domain of an immunoglobulin heavy chain or allelic variant thereof.

26. A polynucleotide encoding a hybrid fusion protein according to claim 25.

5

27. A DNA sequence encoding a polypeptide having a biological property specific for AFM and selected from the group consisting of:

- (a) the DNA sequence set out in FIG 1;
- (b) a DNA which hybridizes under stringent 10 conditions to the DNA of (a); and
- (c) a DNA sequence which, but for the redundancy of the genetic code, would hybridize under stringent conditions to a DNA sequence of (a) or (b).

FIG. I A

CCCCGAGTCTCTGCGCCTCACATAGTGTACAGGACTAAAGCAAATTGATCCAGGGGG 60
 AACACTGTAGACCGTGATATAAAAACACTCTATAAACTGCAATGCTCAATTCTTAGTA 120
 TAACTATTGTTGTTGATATTGATATTATTAGTATTGGTGCTCACAAAAGAGTCTAAATT 180
 CCATAAGTCTTATATTCAAGGCTACTCTTATTTGAAAACTCATTTCTATCACCTTT 240
 TTCTATTTACTCCATATTGAGGCCTCATAAATCCAATTTCATTTCTTGTAA 300

 m k l l k l t g f i f f l f. f -21
 ATGTGGTTCTACAAAGATGAAACTACTAAAACCTACAGGTTTATTTCATTTCTTGTAA 360
 *
 l t e s l t L P T Q P R D I E N F N S T 14
 TTTGACTGAATCCCTAACCCCTGCCACACAAACCTCGGGATATAGAGAACTTCATAGTA 420

 Q K F I E D N I E Y I T I I A F A Q Y V 34
 CTCAAAAATTATAGAAGATAATATTGAATACATCACCATCATTGCATTGCTCAGTATG 480

 Q E A T F E E M E K L V K D M V E Y K D 54
 TTCAGGAAGCAACCTTGAAGAAATGGAAAAGCTGGTGAAAGACATGGTAGAATACAAAG 540

 R C M A D K T L P E C S K L P N N V L Q 74
 ACAGATGTATGGCTGACAAGACGCTCCCAGAGTGTCAAAATTACCTAATAATGTTTAC 600
 *
 E K I C A M E G L P Q K H N F S H C C S 94
 AGGAAAAAAATATGTGCTATGGAGGGGCTGCCACAAAGCATAATTCTCACACTGCTGCA 660

 K V D A Q R R L C F F Y N K K S D V G F 114
 GTAAGGTTGATGCTCAAAGAAGACTCTGTTCTTCTATAACAAGAAATCTGATGTGGGAT 720

 L P P F P T L D P E E K C Q A Y E S N R 134
 TTCTGCCTCCTTCCCTACCCCTGGATCCCGAAGAGAAATGCCAGGCTATGAAAGTAACA 780

 E S L L N H F L Y E V A R R N P F V F A 154
 GAGAATCCCTTTAAATCACTTTATATGAAGTTGCCAGAAGGAACCCATTGTCTTCG 840

 P T L L T V A V H F E E V A K S C C E E 174
 CCCCTACACTCTAACTGTTGCTGTCATTGAGGAGGTGGCCAAATCATGTTGTGAAG 900

 Q N K V N C L Q T R A I P V T Q Y L K A 194
 AACAAAACAAAGTCAACTGCCTTCAAACAAGGGCAATACCTGTACACAAATATTAAAAG 960

 F S S Y Q K H V C G A L L K F G T K V V 214
 CATTTCCTTCTTATCAAAAACATGTCTGTGGGGCACTTTGAAATTGGAACCAAAGTTG 1020

 H F I Y I A I L S Q K F P K I E F K E L 234
 TACACTTTATATATTGCGATACTCAGTCAAAATCCCCAAGATTGAATTAAAGGAGC 1080

 I S L V E D V S S N Y D G C C E G D V V 254
 TTATTTCTCTGTAGAAGATGTTCTTCAAACATGATGGATGCTGTGAAGGGGATGTTG 1140

FIG. 1B

Q C I R D T S K V M N H I C S K Q D S I	274
TGCAGTGCATCCGTGACACGAGCAAGGTTATGAACCATAATTGTTCAAAACAAGATTCTA	1200
S S K I K E C C E K K I P E R G Q C I I	294
TCTCCAGCAAAATCAAAGAGTGCTGTGAAAAGAAAATACCAGAGCGCGGCCAGTGCATAA	1260
N S N K D D R P K D L S L R E G K F T D	314
TTAACTCAAACAAAGATGATAGACCAAAGGATTATCTCTAAGAGAAGGAAAATTTACTG	1320
S E N V C Q E R D A D P D T F F A K F T	334
ACAGTGAAAATGTGTCAAGAACGAGATGCTGACCCAGACACCTTCTTGCAGTTA	1380
F E Y S R R H P D L S I P E L L R I V Q	354
CTTTGAATACTCAAGGAGACATCCAGACCTGTCTATACCAGAGCTTTAAGAATTGTC	1440
I Y K D L L R N C C N T E N P P G C Y R	374
AAATATAACAAAGATCTCCTGAGAAATTGCTGCAACACAGAAAACCCCTCAGGTTGTTACC	1500
*	
Y A E D K F N E T T E K S L K M V Q Q E	394
GTTACGCCGAAGACAAATTCAATGAGACAACTGAGAAAAGCCTCAAGATGGTACAACAAG	1560
C K H F Q N L G K D G L K Y H Y L I R L	414
AATGTAAACATTTCCAGAATTGGGGAGGATGGTTGAAATACCATTACCTCATCAGGC	1620
T K I A P Q L S T E E L V S L G E K M V	434
TCACGAAGATAGCTCCCCACTCTCCACTGAAGAACTGGTGTCTTGGCGAGAAAATGG	1680
T A F T T C C T L S E E F A C V D N L A	454
TGACAGCTTCACTACTTGCTGTACGCTAAGTGAAGAGTTGCCTGTGTTGATAATTGG	1740
*	
D L V F G E L C G V N E N R T I N P A V	474
CAGATTAGTTGGAGAGTTATGTGGAGTAAATCGAACTATCAACCCCTGCTG	1800
D H C C K T N F A F R R P C F E S L K A	494
TGGACCACTGCTGTAAAACAAACTTGCCTTCAGAAGGCCCTGCTTGAGAGTTGAAAG	1860
D K T Y V P P P F S Q D L F T F H A D M	514
CTGATAAAACATATGTGCCTCACCTTCTCAAGATTACCTTCACGCCAGACA	1920
C Q S Q N E E L Q R K T D R F L V N L V	534
TGTGTCAATCTCAGAATGAGGAGCTTCAGAGGAAGACAGACAGGTTCTGTCAACTAG	1980
K L K H E L T D E E L Q S L F T N F A N	554
TGAAGCTGAAGCATGAACACTCACAGATGAAGAGCTGCAGTCTTGTACAAATTGCAA	2040
V V D K C C K A E S P E V C F N E E S P	574
ATGTAGTGGATAAGTGCCTGCAAAGCAGAGAGTCCTGAAGTCTGCTTAATGAAGAGAGTC	2100

3/12

FIG. 1C

K I G N	578
CAAAAATTGGCAACTGAAGCCAGCTGCTGGAGATATGTAAAGAAAAAGCACCAAAGGGA	2160
AGGCTTCCTATCTGTGTGGTGATGAATCGCATTCTGAGAACAAAATAAAAGGATTTT	2220
CTGTAACTGTCACCTGAAATAATACATTGCAGCAAGCAATAAACACACATTGTAAAG	2280
TTAAAAAA	2287

4 / 12

5 / 12

2- 24 2 EIGEN

FIG. 2B

		<u>% Similarity</u>		
		<u>AFM</u>	<u>AFP</u>	<u>ALB</u>
<u>% Identity</u>	<u>AFM</u>		60.4	54.8
	<u>AFP</u>	39.6		59.1
	<u>ALB</u>	35.6	40.2	
	<u>VDB</u>	20.7	21.2	24.4

FIG. 3

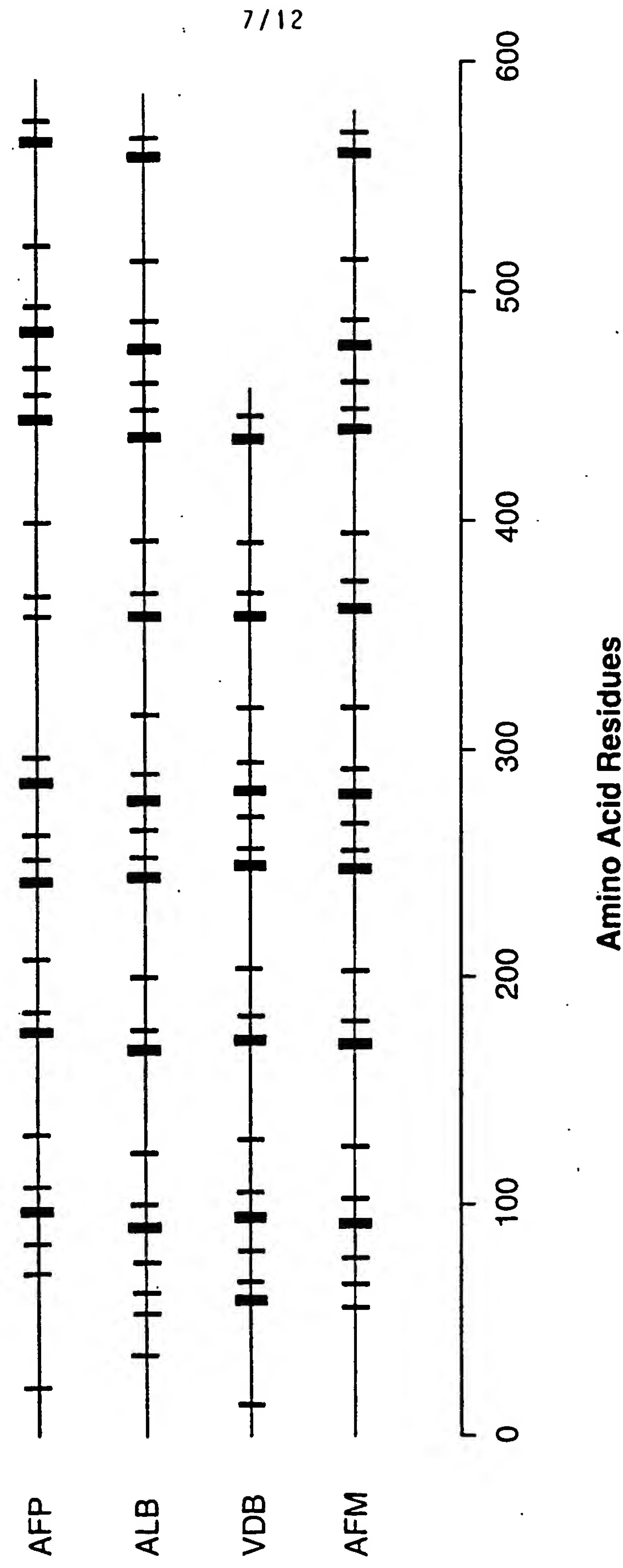


FIG. 4A

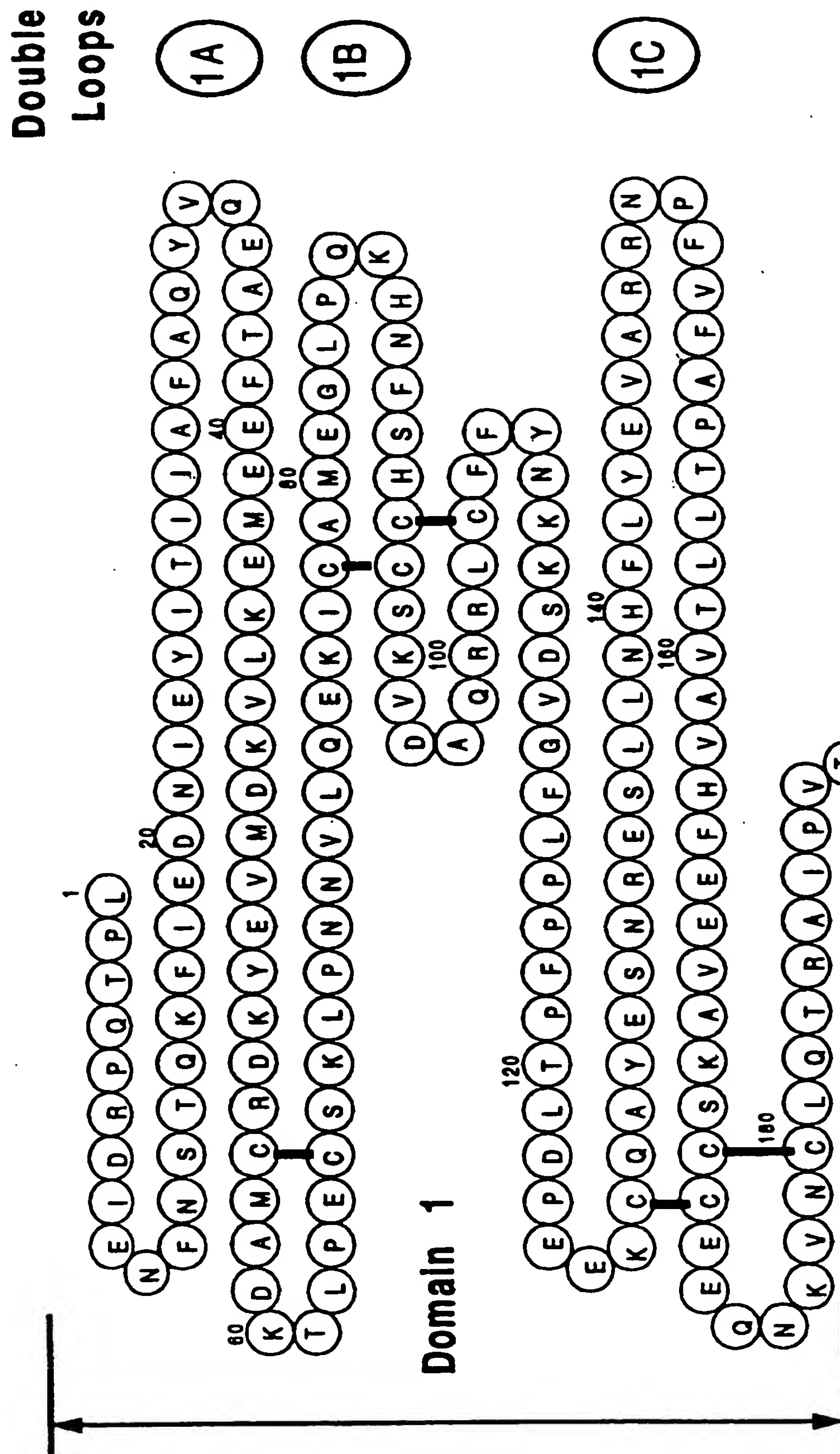


FIG. 4B

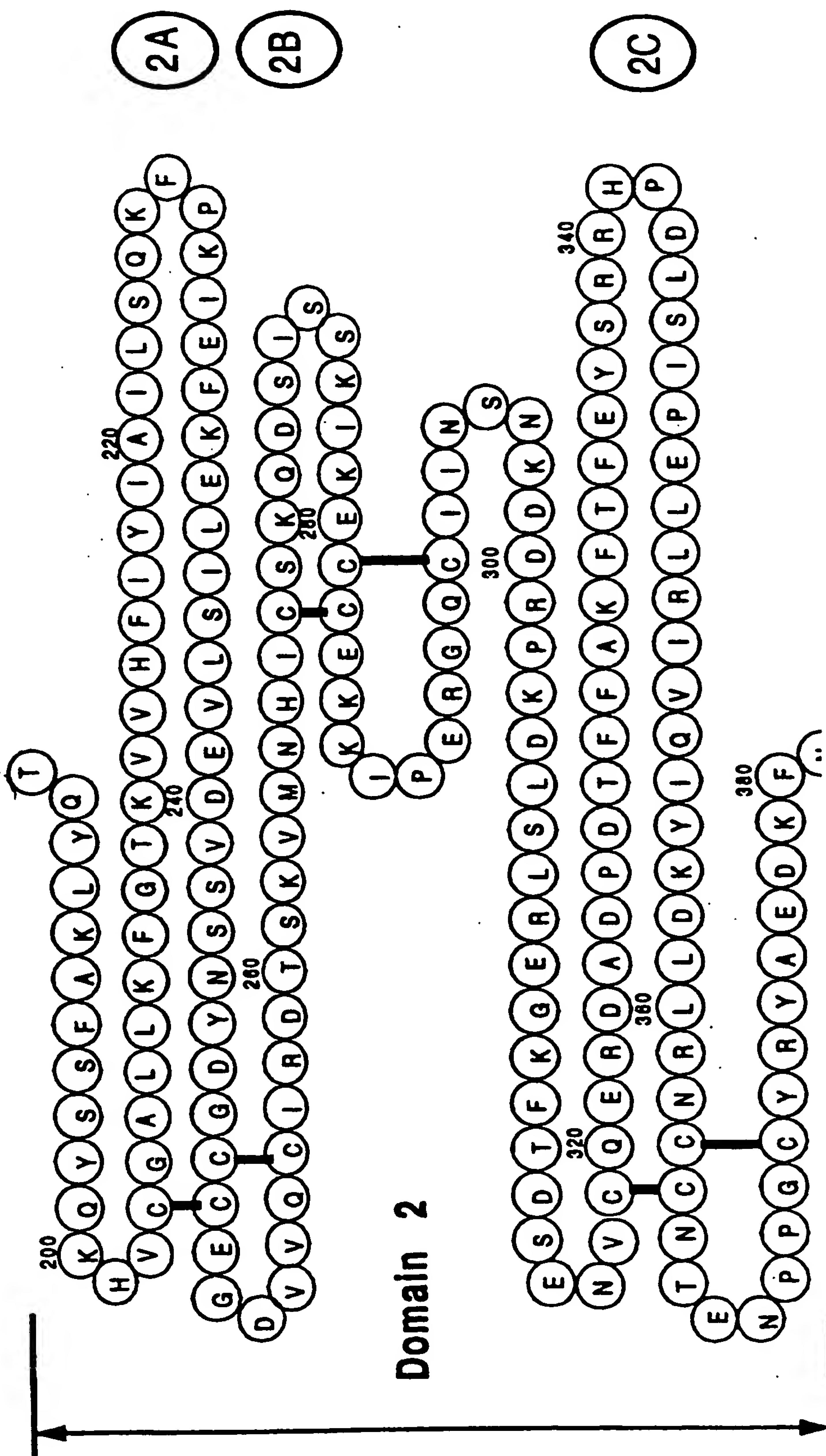
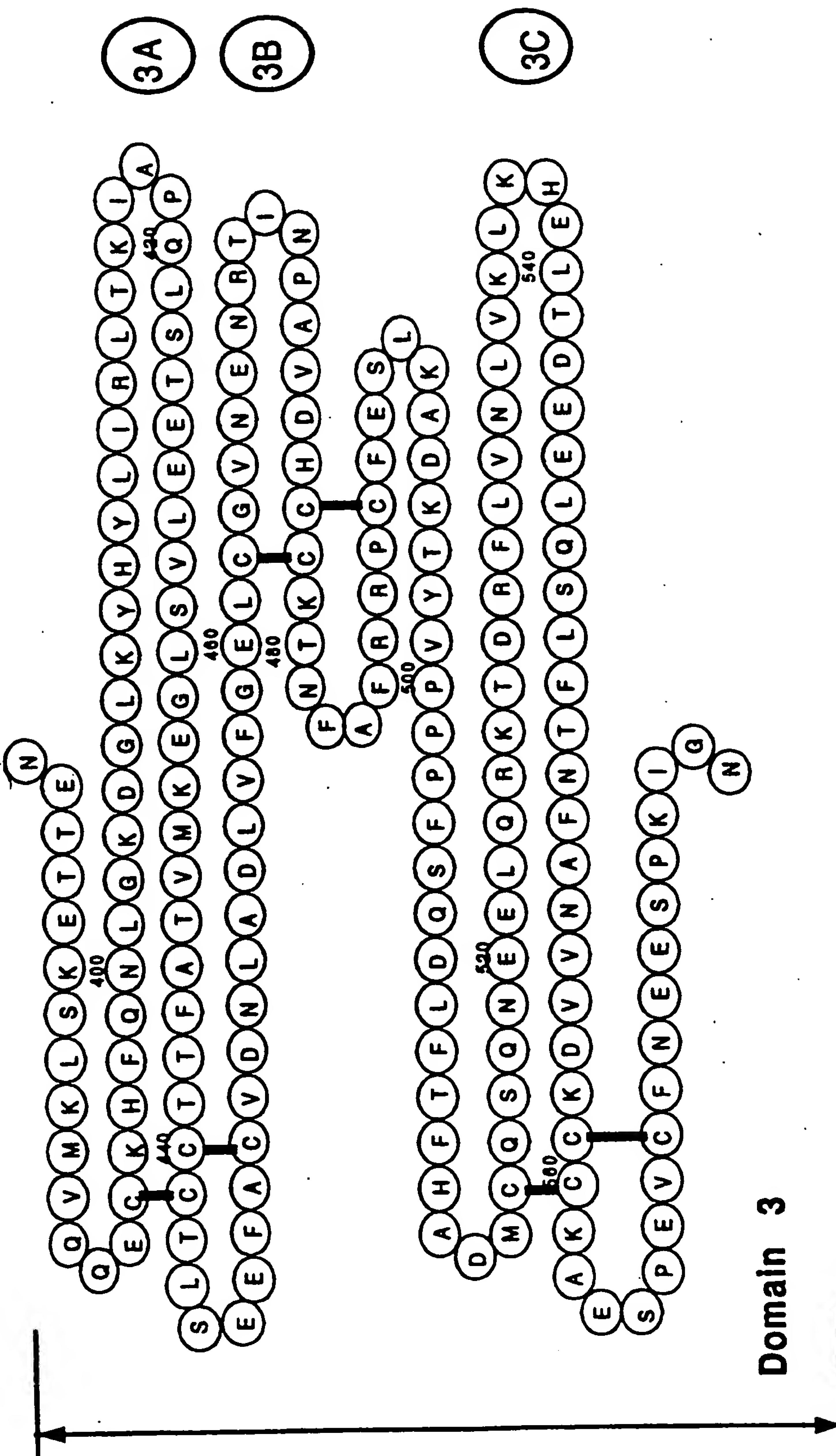
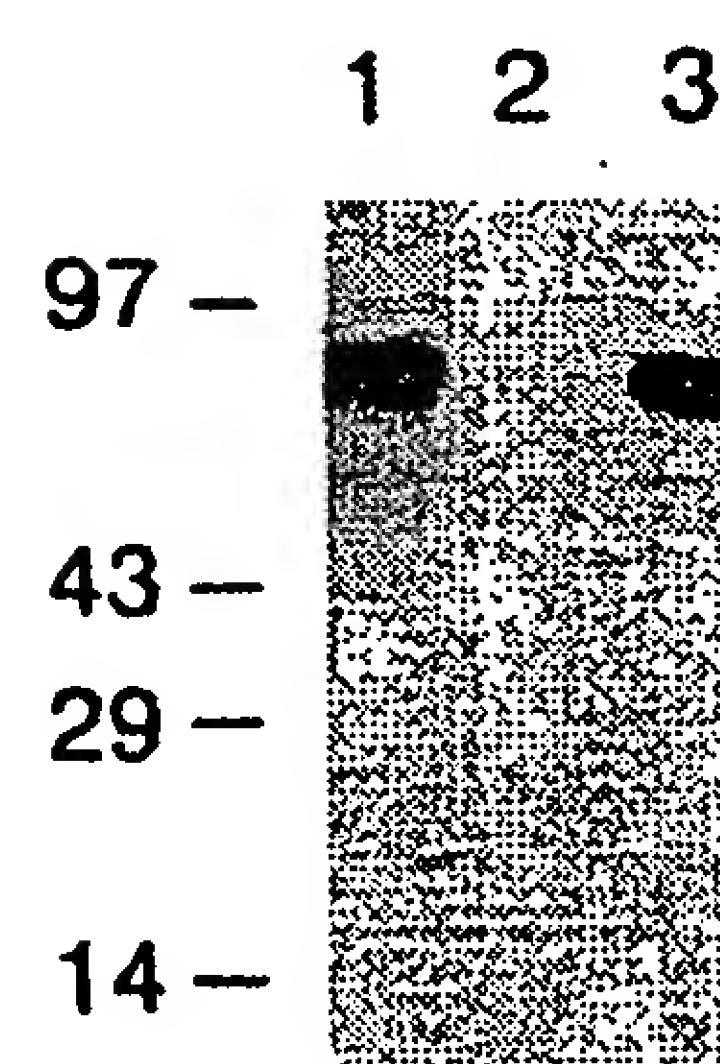


FIG. 4C



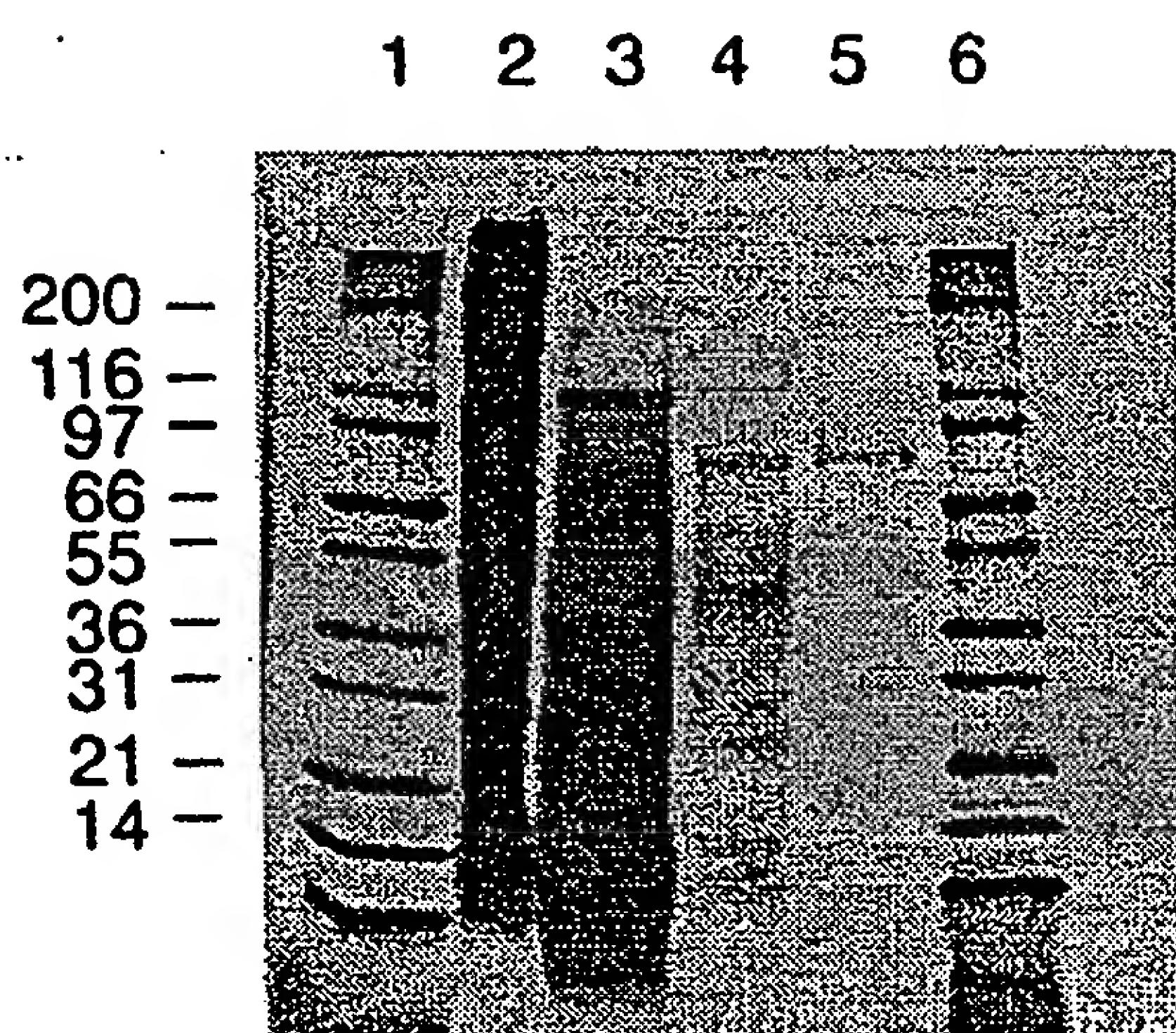
11/12

FIG. 5



12/12

FIG. 6



A. CLASSIFICATION OF SUBJECT MATTER
 IPC 6 C12N15/12 C12N15/62 C07K14/47 C07K16/18 G01N33/53

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	THE JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 269, no. 8, 25 February 1994 pages 5481-5484, BÉLANGER ET AL 'NEW ALBUMIN GENE 3' ADJACENT TO THE ALPHA1-FETOPROTEIN LOCUS' see the whole document ---	1-27
P,X	THE JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 269, no. 27, 8 July 1994 pages 18149-18154, LICHENSTEIN ET AL 'AFAMIN IS A NEW MEMBER OF THE ALBUMIN, ALPHA-FETOPROTEIN, AND VITAMIN D-BINDING PROTEIN GENE FAMILY' see the whole document ---	1-27 -/-



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1

Date of the actual completion of the international search

2 August 1995

Date of mailing of the international search report

10.08.95

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Authorized officer

Sitch, W

C(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
E	WO,A,95 11308 (AMGEN INC) 27 April 1995 see page 6, line 20 - page 35 see SEQ ID NOS: 23 and 24 see page 41 - page 43; example 4 -----	1-14,16, 27

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-9511308	27-04-95	AU-B- 7979194	08-05-95

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